Stereoselective Preparation of 3-Amino-2-fluoro Carboxylic Acid Derivatives, and Their Incorporation in Tetrahydropyrimidin-4(1H)-ones, and in Open-Chain and Cyclic β -Peptides

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The preparation of (2S,3S)- and (2R,3S)-2-fluoro and of (3S)-2,2-difluoro-3-amino carboxylic acid derivatives, $\mathbf{1}-\mathbf{3}$, from alanine, valine, leucine, threonine, and β^3 h-alanine (*Schemes 1* and 2, *Table*) is described. The stereochemical course of (diethylamino)sulfur trifluoride (DAST) reactions with N,N-dibenzyl-2-amino-3-hydroxy and 3-amino-2-hydroxy carboxylic acid esters is discussed (*Fig. 1*). The fluoro- β -amino acid residues have been incorporated into pyrimidinones ($\mathbf{11}-\mathbf{13}$; *Fig. 2*) and into cyclic β -tri- and β -tetrapeptides $\mathbf{17}-\mathbf{19}$ and $\mathbf{21}-\mathbf{23}$ (*Scheme 3*) with rigid skeletons, so that reliable structural data (bond lengths, bond angles, and *Karplus* parameters) can be obtained. β -Hexapeptides Boc[(2S)- β^3 hXaa(αF)] $_6$ OBn and Boc[β^3 hXaa($\alpha,\alpha F$)] $_6$ OBn, $\mathbf{24}-\mathbf{26}$, with the side chains of Ala, Val, and Leu, have been synthesized (*Scheme 4*), and their CD spectra (*Fig. 3*) are discussed. Most compounds and many intermediates are fully characterized by IR- and α -1H-, α -1SC- and α -1F-NMR spectroscopy, by MS spectrometry, and by elemental analyses, [α] $_0$ and melting-point values.

1. Introduction. – The effect of backbone substitution **A** by F-atom(s) (or other heteroatoms) of a peptide consisting of proteinogenic α -amino acids cannot be studied due to hydrolytic instability⁴). In constrast, F-substituted β - and γ -amino acid derivatives **B** [1] [3–6] and **C** [7] are stable. For incorporation of 3-amino-2-fluoro and 3-amino-2,2-difluoro acid moieties into β -peptides **D**-F and **D**-F₂, or for syntheses of β -peptides **E**-F and **E**-F₂ bearing F-atom in each residue, we needed an access to configurationally pure building blocks of type **1**, **2**, and **3** with *N*-Boc protection for solution synthesis [4] and *N*-Fmoc protection for solid-phase peptide synthesis [5].

One aim of the present paper is to describe the preparation of such α -fluoro- β -amino acid derivatives in full detail⁵), and to discuss some mechanistic aspects. A

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⁴⁾ See the discussion in [1]. For a γ , γ -difluoro- δ -amino acid, see [2].

⁵⁾ Only the preparation of the alanine-derived compounds 1-3, R¹=Me, has been described in full detail [4]. For the NMR-structural analyses of four peptides with a central fluoro- or difluoro-substituted building block, see: [1][5][6]. Polyfluorinated peptides of type E-F and E-F₂ are mentioned in a preliminary communication [3], and in the hitherto unpublished master thesis of C. Noti (see Footnote 3). For determination of proteolytic stabilities of F-substituted β-peptides, see [4].

second purpose is to present cyclic and macrocyclic F-substituted β -amino acid derivatives **F**-F and **F**-F₂, which fulfill the following stringent requirements: i) they should be crystalline (for X-ray analysis) and ii) they are likely to have the same conformation in solution (for NMR analysis) and in the solid state; this would provide experimental data (*Karplus* parameters) for the interpretation of NMR spectra and for comparison with structures calculated by *ab initio* or molecular-dynamics methods⁶).

⁶⁾ It turned out that both the NMR analysis and the MD calculation of F-substituted β -peptides gave confusing results, due to lack of reliable parameters [5][6][8].

2. Preparation of the F-Substituted β -Amino Acid Derivatives 1–3. – We have chosen three stereospecific methods for the introduction of F-substituents, all starting from natural α -amino acids alanine, valine, leucine, and threonine⁷). The F-atoms were introduced nucleophilically with (diethylamino)sulfur trifluoride (DAST; 'F-') [9] and electrophilically with (PhSO₂)₂NF ('F+') [9b][10].

The 'work-horse' route (*Scheme 1*) started from the aldehydes $4\mathbf{a} - 4\mathbf{c}$ [11] obtained by N,N-dibenzylation and CO_2H reduction of the corresponding amino acids. *Reetz*'s diastereoselective non-chelation-controlled (BF₃) and chelation-controlled (TiCl₄) cyanohydrin reaction with Me₃SiCN ($\rightarrow 5\mathbf{a} - 5\mathbf{c}$) [12] was followed by the *Pinner* reaction with aqueous workup to give the 3-amino-2-hydroxy carboxyclic acid esters $6\mathbf{a} - 6\mathbf{c}$, treatment of which with DAST led to mixtures of the constitutional isomers 7 and iso-7 with higher regioselectivities in the *l*-series of compounds (*i.e.*, $6 \rightarrow 7$) than in the *u*-series (*i.e.*, $epi-6 \rightarrow epi-7$; see mechanistic discussion in *Sect. 3*). For the preparation of the geminal difluoro- β -amino acid esters, a 'cheaper' non-diastereoselective version of the cyanohydrin reaction was used [13] (*Scheme 1*; lower part). *Swern* oxidation of the mixture of diastereoisomers 6/epi-6 to the α -keto esters and *in situ* treatment with DAST provided the difluoro esters 8 in enantiomerically pure form⁸).

For preparation of the butanoate derivative epi-**7a**, there is a shorter route ($Scheme\ 2,a$) [15]: the N,N-dibenzylthreonine benzyl ester (**9**) undergoes a fluorinating rearrangement when treated with DAST to give epi-**7a**Bn of (2R,3S)-configuration as the major product. As with the other mixtures of isomers **7**/iso-**7** ($Scheme\ 1$, upper part) chromatographic separation from the 'direct' substitution product iso-epi-**7a**Bn and isolation of the pure α -fluoro- β -amino acid ester epi-**7a**Bn was straightforward.

Finally, the methyl Boc- and Cbz-(S,S)-3-amino-2-fluorobutanoates, **1abb** and **1acb**, are accessible by direct fluorination [16] of the corresponding doubly lithiated [17] β ³hAla derivatives with (PhSO₂)₂NF (*Scheme 2*,b).

Having established the preparation of the 3-amino-N,N-dibenzyl-2-fluoro- and -2,2-difluoro-alkanoates **7** and **8**, only simple functional-group manipulations were needed to arrive at the desired building blocks **1**–**3** for peptide synthesis. The reagents and solvents used are collected in the *Table*, together with the starting materials and products of the various conversions. The absolute configurations of compounds **1**–**3** follow from the use of (S)- or L- α -amino acid starting materials, the relative configurations are deduced from an X-ray crystal structure (bottom part of the *Table*), from NMR data and by analogy⁹).

⁷⁾ Since these amino acids are all available in both enantiomeric forms, the enantiomers of the compounds reported herein will be accessible by exactly the same methodology.

⁸⁾ Purification of the keto esters by chromatography turned out to lead to partial racemization. The *in situ* procedure led to the *N*-Boc-amino-difluoro carboxylic acids **3aba**, **3bba**, and **3cba** with er > 97:3, >99:1, and >98:2, respectively, as derived from HPLC on chiral column material of precursor esters. For previous preparations of α,α-difluoro-β-amino acid derivatives, see [14a,b]. For a comprehensive review on the methods of synthesis of geminal difluoro methylene compounds, see [14c].

⁹⁾ The NMR-structural analyses [1] [5] [6] of β-hepta- and β-tridecapeptides of type **D**-F and **D**-F₂ with central F- or F₂-β-amino acid residues **1**, **2**, and **3** (R¹ = Me, R², Y = (βhXaa)_n) provide further support for the correctness of the *relative* and *absolute* configurational assignment of these building blocks.

Scheme 1. Conversion of the Ala., Val., Leu-Derived (S)-Dibenzylamino Aldehydes 4 to the Mono- and Difluoro-Substituted β-Amino Acid Esters 7 and 8. The OH/F replacement with DAST $(6 \rightarrow 7)$ occurs with retention of configuration. The migratory fluorination, in which the F-atom winds up at C(3) (replacing NBn_2) and the amino group at C(2) (replacing OH), takes place with inversion of configuration on both centers $(6 \rightarrow iso-7)$.

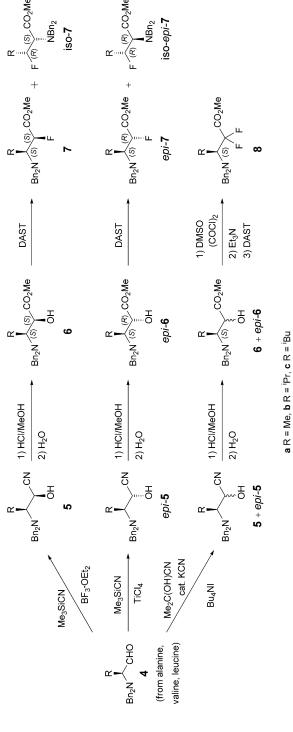
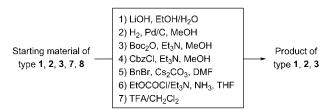
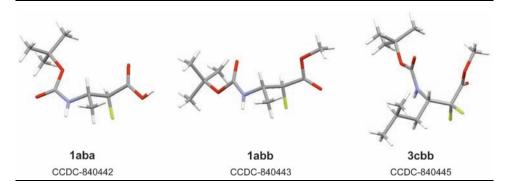


Table. Reagents Used for the Functional-Group Manipulations to Convert the Primary Products 7 and 8 of Fluorination to the Starting Materials 1–3. Interconversions of compounds of type 1 and 2 are also included in this Table. For the specification with the letters a, b, c, d, see the Formulae in the Introduction. The X-ray data for compounds 1aba, 1abb, and 3cbb have been deposited with the Cambridge Crystallographic Data Centre.



Starting material	Reaction conditions	Product	Starting material	Reaction conditions	Product
7a	1, 2, 3	1aba	1abb	1	1aba
7b	1, 2, 3	1bba	1acb	1	1aca
7c	1, 2, 3	1cba	1cba	5	1cbc
epi- 7a	1, 2, 3	2aba	2cba	5	2cbc
epi- 7a Bn	2, 3	2aba	3cba	5	3cbc
epi- 7a Bn	2, 4	2aca	1aca	6, 2	1aad
epi- 7b	1, 2, 3	2bba	2aca	6, 2	2aad
ері- 7с	1, 2, 3	2cba	3aca	6, 2	3aad
8a	1, 2, 3	3aba	1cbc	7	1cac
8a	1, 2, 4	3aca	2cbc	7	2cac
8b	2, 3	3bbb	3cbc	7	3cac
8c	2, 3	3cbb	3bbb	1	3bba
	,		3cbb	1	3cba



Before describing the use of the fluorinated β -amino acids for tetrahydropyrimidinone and peptide synthesis, some comments about the mechanistic course of the DAST reaction are appropriate.

3. Stereo- and Regiochemical Course of the DAST Reactions $6 \rightarrow 7$. – As we can see from *Schemes 1* and 2, a, the desired β -amino- α -fluoro-acid esters 7 are formed with

Scheme 2. Alternative Routes to α -Fluoro- β -amino Acid Derivatives. a) Treatment of an N,N-dibenzylthreonine ester with DAST, and b) direct electrophilic fluorination of an N-carbamoyl- β -amino acid ester through a doubly lithiated species.

Me Me Me Me NBn₂
$$(S)$$
 CO_2Bn THF Bn_2N (S) (R) CO_2Bn (F) (R) $($

retention of configuration 10), when we start from the β -amino- α -hydroxy-acid esters 6, and with inversion on both stereogenic centers, when we start from the α -amino- β -hydroxy ester 9. The isomeric α -amino- β -fluoro carboxylic acid esters iso-7 and iso-*epi*-7 are formed with inversion on both centers and with retention on the F-substituted C-atom, respectively. This, at first sight, somewhat confusing stereochemical outcome is caused by the intermediacy of N,N-dibenzylaziridinium ions [15a,b], as outlined in Fig.~1, b-f. Both, the ring closure to, and the ring opening of the three-membered ring occur with an S_N 2-type inversion of configuration, and the regiochemical course is determined by the relative rates of C-N bond cleavage next to the ester group and next to the substituent R on the three-membered ring. As can be seen from the table in Fig.~1, the carbonyl-assisted opening of the aziridinium ring is preferred in the *trans*-substituted series, while there is a delicate dependence on the type of substituent in the *cis*-series of aziridinium intermediates. Concomitantly, the yields of the desired β -amino- α -fluoro-acid derivatives of (2R,3S)-configuration (*epi*-7) are lower than those of the (S,S)-isomers 7.

4. Tetrahydropyrimidin-4(1H)ones and Cyclo- β -peptides Containing the F-Substituded Amino Acid Residues of 1a, 2a, and 3a. – As mentioned in the *Introduction*, there is lack of structural and NMR data for F-substituted carbonyl compounds. To correlate X-ray-crystal solid-state structures with NMR-solution structures, the compounds should have rigid skeletons. Thus, we turned to previous work on non-fluorinated β -amino acid derivatives, which had been shown to have a high degree of crystallinity, and which had structures that are likely not to be subject to dynamic conformational equilibrations on

$$Y^{Re}$$
- $C_{R_2}^{R_3}$ retention X^{Re} - $C_{R_2}^{R_3}$ inversion R^3 R^2

¹⁰⁾ Note that the CIP convention may not be useful for assigning retention or inversion of configuration because priority orders may change. A useful test is to assign whether the new substituent is located in the same or in the opposite half-space of the stereogenic center as compared to the replaced substituent:

a)
$$Bn_2N$$
 CO_2R' Bn_2 B

Fig. 1. Steric and regiochemical course of the DAST reactions with dibenzylamino hydroxy esters 6 and 9. a) Reaction with DAST converts the OH group to a leaving group, which is not replaced intermolecularly by F^- but intramolecularly by Bn_2N . b) The resulting aziridinium ion can react with F^- in the α -carbonyl (α) or in the β -carbonyl position (β). c) In the trans-substituted aziridinium ring, an S_N2 -assisting conformation of the ester group is possible (high regioselectivity of α attack). d) In the cissubstituted aziridinium ring, there are three substituents on the same face of the three-membered ring, which causes steric hinderence of the S_N2 -assisting ester-group conformation, allowing for competing ring-opening (β) next to the R group (poor regioselectivity (epi-7/iso-epi-7)). e) and f) trans- and cisaziridinium-ion intermediates, relative rates of ring opening, and yields of purified products 7 and epi-7. The high selectivity of formation of epi-7b through as cis-aziridinium ion might be due to a kind of neopentyl effect: a Me group of the iPr-substituent blocks the S_N2 trajectory of the attacking F^- nucleophile, thus favoring the ring opening next to the ester group.

the NMR time-scale. These were hexahydropyrimidin-4-ones **G** [18], and cyclo- β -triand -tetrapeptides **H** [19] and **I**, respectively [20]¹¹) (*Fig.* 2).

Tetrahydropyrimidin-4(1H)-ones **11–13**. Thus, we treated the amino acid amides **1aad**, **2aad**, and **3aad** with pivalaldehyde in refluxing CH_2Cl_2 (with azeotropic removal

These cyclo- β -peptides assemble to stacks in the solid state. A cyclo- β -tripeptide has recently been used to construct a mimic of CD40L with a K_D value of 2.4 nM [21].

Fig. 2. Cyclic derivatives G, H, and I of β -amino acids and F-substituted tetrahydro-pyrimidinones 11-13. The numbers in the formulae of 11-13 are bond distances in Å, determined by X-ray crystal-structure analysis I^3). For I^3 , two sets of bond lengths are given, since there are two different conformations in the crystal unit cell of this compound. Presentation I indicates the I^3 no I^3 interaction facilitating ring opening to the imino amides I^3 and I^3 .

of H_2O), which led to the tetrahydropyrimidin-4(1H)-ones **11**, **12**, and **13**, respectively (Fig. 2). The yields of purified, crystalline products were moderate or low (52, 43, and 16%, resp.), which was, at least partially, due to their lability: when dissolved in a protic solvent such as MeOH, they underwent ring opening to yield imino amides **14** and **15** (NMR-tube experiment). Especially the F_2 -substituted compounds of type **13** are prone to undergo this isomerization. The cis- and trans-isomers **13a** and **13b** co-crystallized in a 1:1 ratio and could not be separated. The crystals of **12** contain two conformers in the unit cell. Benzyloxycarbonylation of the tetrahydropyrimidin-4(1H)-

ones 11–13 gave more stable ¹²) derivatives Cbz-11, Cbz-12, and Cbz-13, respectively. X-Ray crystal structures ¹³) of compounds 11–13 exhibit pronounced pyramidalization of the amino N(5) (Δ between 0.30 and 0.44 Å), such that the virtual lone-pair lobe is *antiperiplanar* to the C(6)–N(1) bond, with concomitant bond-length shortening and lengthening. In the Cbz derivatives, the N(5)-atom is, of course, sp²-hybridized, and the bond lengths (in Cbz-12) are more or less normal ¹⁴). The instability of tetrahydropyrimidin-4(1*H*)-ones is clearly caused by a stereoelectronic effect ($n_N \rightarrow \sigma^*(C-N)$), as indicated by the presentation **J** in *Fig.* 2.

Cyclo-β-tripeptides **17–19**. The synthesis was accomplished according to traditional protocols (*Scheme 3*). Coupling of the *N*-Boc-fluoro-amino acids with the dipeptide ester H- β Gly- β Gly-OMe using HBTU/NMM¹⁵) furnished the *N*-Boc- β -tripeptides **16a–16c** in high yields. There are numerous approaches for the cyclization of peptides, many with miserable yields. The route *via* pentafluorophenyl esters, as developed by *U. Schmidt et al.* [24], was the method of choice, as it had been successfully employed for β -peptides before [19b,d]: transesterification of **16a–16c** to the active esters, Boc-deprotection, and slow addition (*via* syringe pump) of solutions of the resulting TFA salts of the tripeptide pentafluorophenyl esters to a dilute (3.3 mM) solution of *Hünig* base (DIPEA in MeCN, 70°) gave the cyclo- β -tripeptides **17–19** in good overall yields (*Scheme 3*, left). Like the non-fluorinated analogs [19b,c], the cyclization products are insoluble in essentially all common solvents and precipitated from the hot, dilute reaction mixture, to be isolated by simple filtration. NMR Spectra were recorded in a mixture of CDCl₃ and TFA.

Cyclo-β-tetrapeptides 21 – 23. In an attempt to prepare cyclo-β-dipeptides consisting of one F-substituted residue and one β-Gly moiety, we prepared the N-Boc-protected peptide methyl esters 20a-20c. By the same procedure as for the tripeptides, the dipeptide pentafluoro-phenyl esters were prepared, and their solutions were added to the $H\ddot{u}nig$ -base solution (MeCN, 70°). To our surprise 16), the cyclo-β-tetrapeptides

¹²⁾ The F₂-substituted Cbz-13 was still quite unstable and could be isolated in only 6% yield as an oil (see Exper. Part). We had noticed the instability of such hydro-pyrimidinones with non-F-substituted derivatives before [18f].

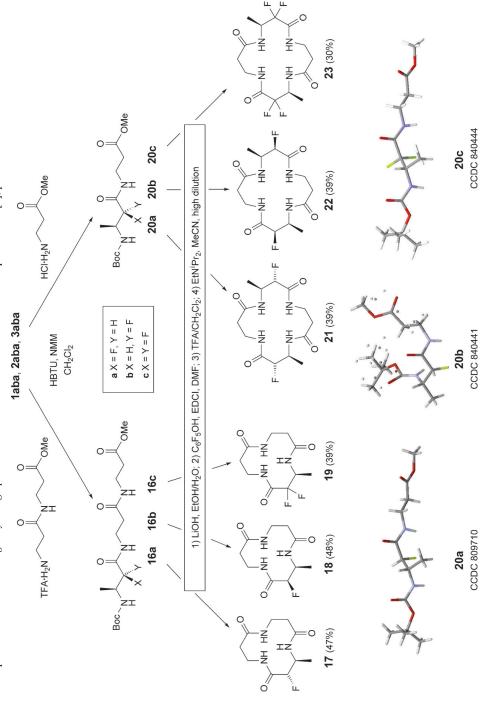
¹³⁾ The crystal structures of the heterocycles 11-13 of type G and of the cyclo-β-tri- and -tetrapeptides H and I will be published separately, together with detailed NMR analyses and ab initio calculations [22].

¹⁴⁾ The dihydrobenzopyrimidin-1(1H)-ones of type i are also somewhat unstable. In crystal structures, their N(1)-atom is, however, only slightly or not at all pyramidalized (π-conjugation is stronger than n → σ* interaction) [23].

¹⁵⁾ For abbreviations not specified in the Schemes, Table, and Figures, see introduction of the Exper.
Part

¹⁶⁾ A 'reverse' case was reported by Vilarrasa and co-workers, who tried to prepare a cyclo-β-tetrapeptide under the same conditions and were disappointed to obtain the eight-membered ring of a cyclo-β-dipeptide [20b].

20c. Overall yields of conversions from the open-chain N-Boc-protected peptide esters 16 and 20 are given. The X-ray data for compounds 20b and 20c have been deposited with the Cambridge Crystallographic Data Centre. The structure of 20a has been published in [6]; parts of the structure of 20b are disordered. Scheme 3. Synthesis of F-Substituted Cyclo- β -tripeptides 17–19 and Cyclo- β -tetrapeptides 21–23, and X-Ray Crystal Structures of the Linear Precursors 20a–



21–23 were isolated in acceptable yields (30–40%; *Scheme 3*, right). Again, these cyclic peptides have high melting points and poor solubilities in common organic solvents, like the non-fluorinated counterparts [19d][20c], and as with those, we were not able to prepare suitable single crystals for conventional X-ray analyses.

There are ongoing attempts to determine the structures of some of the F-substituted cyclo- β -peptides 17 – 19 and 21 – 23 by powder-diffraction techniques¹³), a method that had been successful with the simple $(\beta^3 hAla)_4$ derivatives [20c].

5. Synthesis and Spectroscopic Characterization of β -Hexapeptides 24 – 26 with F-Substitution in Each Residue. – From the NMR analyses of β -peptides containing a single, central fluoro- or difluoro- β -amino acid residue 1, 2, or 3 [1][5][6], we would expect that a β -hexapeptide 24b, built of (S,S)-2-fluoro- β hVal, - β hAla, and - β hLeu residues does not fold to an (M)- β ₁₄-helix [25]. The isomeric hexapeptide 25b with the corresponding (S,S)-fluoro-amino acid building blocks, on the other hand, could fold to an (S)-helix, with axial disposition [25] of an F-atom in each residue. For the hexapeptide 26b, with two geminal F-atoms in each amino acid moiety of the (S)-hAla-S-hLeu)₂ chain, a weaker tendency for folding to the S₁₄-helix must be envisaged [6].

Synthesis. The assembly of the three F-substituted β -hexapeptides 24b-26b was performed as outlined in Scheme 4, by using a classical solution-phase Boc strategy for the coupling (with NMM/HOBt/EDC or NMM/HATU¹⁵), and a C-terminal benzyl ester group. After attaching the β hAla and β hVal units to the β hLeu-benzyl esters (*i.e.*, **1cac**, **2cac**, **3cac**), the tripeptide derivatives **25abb-26abb** were divided into two portions, one to be Boc-deprotected (TFA in CH₂Cl₂), the other one to be debenzylated (H₂/Pd-C in MeOH).

Fragment coupling of the two trimers gave the terminally protected hexapeptides **24bbb** – **26bbb**, which were investigated as such or after deprotection to **24baa** – **26baa**, respectively. Most intermediates and final products of these syntheses were characterized by melting points, $[\alpha]_D$ values, 1H -, $^1^3C$ -, $^1^9F$ -NMR, MS, IR, CD spectroscopy, and elemental analyses (see *Exper. Part*).

CD and NMR Spectra. In our work on β -peptides, we have preferentially used MeOH as solvent for CD and NMR recordings; it turned out that some of the fluorohexapeptide derivatives are poorly soluble in this solvent. For the high dilution (0.2 nm) of solutions for recording CD spectra, the solubility in MeOH was sufficient, but for NMR recordings, we had to turn to more polar solvents such as (D₆)DMSO (H-bond destroying) or CF₃CD₂OD (TFE, favoring secondary-structure formation).

The *CD spectra* of the hexapeptide **24bbb**, consisting of *like*-fluoro- β -amino acid residues in hexafluoro-isopropanol ((CF₃)₂CHOH; HFⁱPrOH), of the *unlike*-isomer **25bbb** and its deprotected form **25baa** (in MeOH, TFE, and HFⁱPrOH), and of the hexapeptide **26bbb** with geminal difluoro groups and its unprotected form **26baa** (in the same solvents) are shown in *Fig. 3, a, b,* and *c*, respectively. The peptide consisting of (*S,S*)-building blocks exhibits two strong *Cotton* effects (θ ca. 80,000) at 213 (positive) and 195 nm (negative) in HFⁱPrOH; due to lack of comparison, it is impossible to interpret this spectrum. If we compare the CD spectra of the hexapeptide derivatives with one F-atom per amino acid unit and (2*R*,3*S*)-configuration (*Fig. 3,b*) and with two F-atoms per amino acid unit (*Fig. 3,c*) in the standard solvent MeOH, we would

Scheme 4. Synthesis of the β -Tri- and β -Hexapeptide Derivatives **24a** – **26a** and **24b** – **26b**, Respectively, with One or Two F-Sustituents in Each Amino Acid Residue. The syntheses were carried out by coupling in solution using the Boc strategy. For details, see the Exper. Part.

conclude that there is no secondary structure in the first case (weak extrema) and an unknown structure in the second case (strong maxima of θ up to 100,000 near 220 and 215 nm)¹⁷). There are two surprises in the CD spectra.

i) In the fluorinated alcohols as solvents, the unprotected β -hexapeptide **25baa** with (R,S)-building blocks exhibits a CD pattern, which 'the experts in the area' would associate with a left-handed β_{14} -helix (minimum, albeit weak, near 215, and intensive maximum near 195 nm; Fig. 3,b); such a helix would have all F-atoms in the thermodynamically more stable axial disposition (with antiperiplanar arrangement of F- and carbonyl O-atom, i.e., F-C-C=O dihedral angle of 180° [6]). The terminally protected form **25bbb** has a more or less flat CD pattern near zero in all three solvents (Fig.3,b).

¹⁷) A typical CD spectrum of a β -peptide folding to a 3_{14} -helix shows a strong *negative Cotton* effect near 215 nm [25].

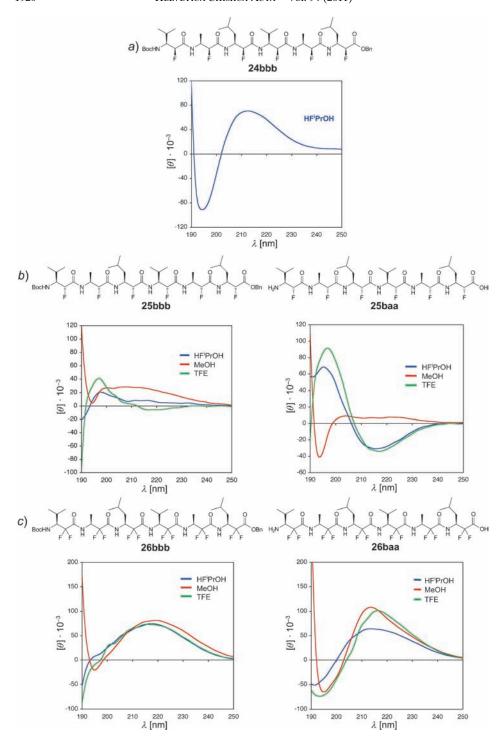


Fig. 3. CD Spectra of the β -hexapeptide derivatives with one or two F substituents in each residue

ii) The β -hexapeptides with geminal difluoro substitution in each residue give rise to CD spectra, which are almost superimposable with those for the protected form **26bbb** (Fig. 3,c), and quite similar to those for the unprotected form **26baa** in all three solvents (Fig. 3,c), meaning that – in first approximation – the secondary-structure-inducing fluorinated solvents do not change the backbone conformation, as compared to MeOH.

Caveat: Suggestions, made in this section, about possible structures of the fluorinated hexapeptides must be taken with due care: we have reiterated that the CD spectra of β -peptides can be deceiving, to say the least [25][26].

NMR Spectra of the hexafluoro- and dodecafluoro- β -hexapeptides with and without terminal protection (*N*-Boc, OBn) were recorded with solutions in CF₃CD₂OD/TFA, CD₃OH, or (D₆)DMSO (see *Fig. 4*); they are described in the *Exper. Part*; an interpretation will have to wait for our detailed NMR-structural analyses with the parameters determined from the cyclic compounds.

6. Conclusions and Outlook. – Configurationally uniform mono- and difluoro-βamino acid derivatives and peptides with and without N- and/or C-terminal protections have been successfully prepared by known methods. The intermediacy of N,N-dibenzylaziridinium ions in the DAST reaction of vicinal amino-hydroxy-substituted C-chains has been confirmed. The regioselectivity of S_N 2-type ring opening of these threemembered rings has been shown to be subject to intriguing, subtle substitution effects. The expected crystallinity of cyclic derivatives of the fluorinated β -amino acids has come true. The tetrahydropyrimidin-4(1H)-ones readily form suitable single crystals, for X-ray structure determination. The cyclic β -tri- and β -tetrapeptides could be purified to give correct elemental analyses, but are poorly soluble, so that we have to resort to - ongoing - powder X-ray diffraction measurements in a synchroton beam. The results of the X-ray-structural investigations will be reported separately, together with full NMR-structural analyses and computational results. The multiply Fsubstituted β -peptides have, so far, resisted all attempts of structure determination, due to poor solubility in common solvents and to lack of reliable Karplus parameters. With the data collected from the cyclic derivatives, we are now in the process of making another attempt. The CD spectra of these linear β -hexapeptides, as reported herein, can be merely considered as 'fingerprints', and the discussed similarities of CD patterns with those of β -peptides of known structure may be accidental.

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Experimental Part

1. General. Abbreviations: β-hAa: β-Homoamino acid, Bn: benzyl, Boc: (tert-butoxy)carbonyl, Boc₂O: di(tert-butyl) dicarbonate, DAST: (diethylamino)sulfur trifluoride, EtNⁱPr₂ (Hünig base): ethyl(diisopropyl)amine, EDC: 1-[3-(dimethylamino)propyl]-3-ethylcarbodiimide hydrochloride, FC: flash chromatography, HATU: O-(7-azabenzotriazol-1-yl)-N,N,N',N'-tetramethyluronium hexafluorophosphate, HFⁱPrOH: 1,1,1,3,3,3-hexafluoropropan-2-ol, HBTU: O-(benzotriazol-1-yl)-N,N,N',N'-tetra-

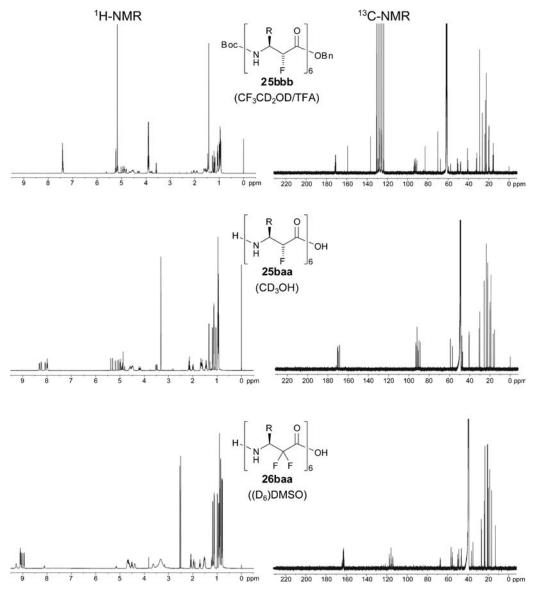


Fig. 4. ¹H- and ¹³C-NMR Spectra of the hexapeptide derivatives **25bbb**, **25baa**, and **26baa**. For ¹H- and ¹⁹F-NMR spectra of **26bbb**, see the *Exper. Part.* For **25bbbb**, ¹H-, ¹³C-, and ¹⁹F-NMR spectra are described in the *Exper. Part.*

methyluronium hexafluorophosphate, HOBt: 1-hydroxy-1H-benzotriazole, h.v.: high vacuum (0.01 – 0.1 Torr), NMM: 4-methylmorpholine, NFSI: N-fluorobenzene-sulfonimide, TFA: trifluoroacetic acid, TFE: 2,2,2-trifluoroethanol.

DMSO and Et₃N were distilled over CaH₂ and stored over 4-Å molecular sieves. Solvents for FC and workup procedures were distilled over Sikkon (anh. CaSO₄; Fluka). α -Amino acids were purchased from

Bachem, Senn, or *Degussa.* Dry THF was distilled from sodium and benzophenone; dry CH_2Cl_2 was distilled from CaH_2 . All moisture-sensitive reactions were carried out under a positive pressure of N_2 or Ar in oven-dried glassware (140°) . All other reagents and solvents were used as received from *Fluka* or *Aldrich.* Sat. HCl/MeOH soln. was prepared by bubbling anh. HCl gas into MeOH at 0° (ice bath). Aldehydes **4b** and **4c** [11], cyanohydrins **5b** and **5c** [12], alanine derivatives **1aba**, **2aba**, **3aba**, **6a**, *epi-6a*, and **7a** [4] were synthesized according to published procedures.

TLC: Merck silica gel 60 F₂₅₄ plates; detection with UV light or by dipping into a soln. of ninhydrin (0.6 g), AcOH (2 ml), H₂O (13 ml), and BuOH (285 ml), or a soln. of phosphomolybdic acid (25 g), $Ce(SO_4)_2 \cdot H_2O(10 g)$, conc. $H_2SO_4(60 ml)$, and $H_2O(940 ml)$, followed by heating. FC: Fluka silica gel 60 (40-63 μm); at ca. 0.3 bar. Anal. RP-HPLC: Knauer HPLC system K 1000, pump type 64, EuroChrom 2000 integration package, degaser, UV detector K 2000 (variable-wavelength monitor), Macherey-Nagel C_8 column (Nucleosil 100-5 C_8 (250 × 4 mm)); at 220 nm. Prep. RP-HPLC: Knauer HPLC system, pump type 64, programmer 50, UV detector (variable-wavelength monitor), Macherey-Nagel C₈ column (Nucleosil 100-7 C₈ (250 × 21 mm)); at 220 nm. M.p.: Büchi 510 apparatus; uncorrected. Optical rotations ($[\alpha]_{\rm in}^{\rm r.t.}$): Perkin-Elmer 241 polarimeter (10 cm, 1-ml cell) at r.t.; the solvent and the concentration (g/100 ml) are given in the procedures. CD Spectra: Jasco J-710 spectrophotometer, recording from 190 to 250 nm at 20°; 1-mm rectangular cell; average of five scans, corrected for the baseline; peptide concentration, 0.2 mm; band width, 1.0 nm; resolution, 0.2 nm; sensitivity, 100 mdeg; response, 0.5 s; speed, 20 nm/min.; molar ellipticity $[\theta]$ in deg cm² mol⁻¹ (λ in nm); smoothing by *Jasco* softwares). IR Spectra: Perkin-Elmer 782 spectrophotometer. NMR spectra: Bruker AMX 600 (1H: 600 and ¹³C: 150.9 MHz), AMX 500 (¹H: 500 and ¹³C: 125 MHz), AMX 400 (¹H: 400 and ¹³C: 100 MHz), or AV-400 (1H: 400, 13C: 100, and 19F: at 376 MHz), or Varian Gemini 300 (1H: 300, 13C: 75, and 19F: 282 MHz); chemical shifts δ in ppm and coupling constants J in Hz. HR-MS: IonSpec Ultima 4.7 (HR-ESI-MS and HR-MALDI-MS) or Bruker Reflex (MALDI-TOF-MS) spectrometer; in m/z (% of basis peak). Elemental analyses: performed in the Microanalytical Laboratory of the Laboratorium für Organische Chemie, ETH Zürich.

2. General Procedures 1–14 (GP 1–14). Fluorination of β -Homoalanine Methyl Esters: GP 1. A soln. of BuLi (1.6M hexane, 2.2 equiv.) was added to a soln. of ${}^{\rm i}{\rm Pr}_2{\rm NH}$ (2.2 equiv.) in THF (0.5M) at -78° under Ar. The mixture was stirred for 1 h at -78° , then a soln. of the N-Boc- or N-Cbz-amino acid ester (1 equiv.) in THF (0.5M) was added. The mixture was stirred for 1 h at -78° and then a soln. of NFSI (2.5 equiv.) in THF (1M) was added dropwise via syringe. The resulting yellow mixture was stirred for 2.5 h at -78° and for 2 h at 0°. Sat. NH₄Cl was added, and the aq. layer was extracted with CH₂Cl₂ (3×). The combined org. phases were dried (MgSO₄), and the solvent was removed *in vacuo*. The crude product was purified by FC.

Fluorination Reactions with DAST: GP 2. All reactions were performed in PET flasks under N_2 . To a soln. of the appropriate starting material (1 equiv.) in CH_2Cl_2 (2 ml/mmol), DAST (1.5–3 equiv.) was slowly added at 0° (ice bath) or r.t. The mixture was stirred at 0° or r.t. for 3–6 h, poured into H_2O , cautiously neutralized by the addition of solid K_2CO_3 , and extracted with Et_2O (2×). The combined org. layers were dried (MgSO₄), filtered, and evaporated. The crude product was purified by FC.

Methyl Ester Hydrolysis: GP 3. A mixture of the methyl ester (1 equiv.) and LiOH \cdot H₂O (3 equiv.) in EtOH/H₂O 2:1 (4 ml/mmol) was well stirred for 1 h at r.t., cooled to 0° (ice bath), and neutralized by the addition of 1m HCl. The mixture was diluted with H₂O and extracted with AcOEt or CH₂Cl₂ (3×). The combined org. layers were dried (MgSO₄), filtered, and evaporated. The crude product was used without further purification.

Benzyl Hydrogenolysis: GP 4. To a soln. of the Bn-protected compound in MeOH (10 ml/mmol), a cat. amount of Pd/C (10%, 30 mg per equiv. of Bn group) was added. The apparatus was evacuated and flushed three times with $\rm H_2$, and the mixture was vigourously stirred for 12-24 h. The catalyst was filtered off through Celite, washed several times with MeOH, and the combined org. layers were evaporated. The crude product was used without further purification.

Boc Protection: GP 5. To a soln. of the amine or the TFA salt (1 equiv.) in MeOH (3 ml/mmol), Boc₂O (1.5 equiv.) and Et₃N (3-5 equiv.) were added. The mixture was stirred at r.t. for 12 h, concentrated under reduced pressure, dissolved in AcOEt and washed successively with 0.5M HCl, sat.

 K_2CO_3 , and brine. The org. layer was dried (MgSO₄), filtered, and evaporated, and the crude product was purified by FC.

Benzyl Ester Formation: GP 6. According to the procedure described in [27], a mixture of the carboxylic acid (1 equiv.) and dry Cs_2CO_3 (1 equiv.) in DMF (3 ml/mmol) was stirred for 10 min at r.t., and treated with BnBr (1.2 equiv.). After stirring for 12 h at r.t., the mixture was diluted with AcOEt and washed successively with 1 MHCl (2×), sat. K_2CO_3 , and brine. The org. layer was dried (MgSO₄), filtered, and evaporated. The crude product was purified by FC.

Non-Stereoselective Preparation of Cyanohydrins: GP 7. In analogy to the procedure described in [13], a vigorously stirred, biphasic soln. of the dibenzylamino aldehyde (1 equiv.) in hexane/ H_2O 3:1 (1.5 ml/mmol) was treated with acetone cyanohydrin (=2-hydroxy-2-methylpropanenitrile; 1.5 equiv.) at r.t. After stirring for 5 min, cat. amounts of KCN (0.03 equiv.) and Bu_4NI (0.01 equiv.) were added. The mixture was stirred at r.t. for 2 h, poured into H_2O , and extracted with Et_2O (3×). The combined org. phases were washed with brine, dried (MgSO₄), filtered, and evaporated. The crude epimeric mixture of cyanohydrins was used without further purification.

Methanolysis of Cyanohydrins: GP 8. A soln. of the cyanohydrin (1 equiv.) in sat. HCl/MeOH (5–10 ml/mmol) was stirred at r.t. for 12 h, concentrated under reduced pressure, poured into H_2O , cautiously neutralized by the addition of solid K_2CO_3 , and extracted with AcOEt (3×). The combined org. layers were dried (MgSO₄), filtered, and evaporated. The crude product was purified by FC.

Oxidation of α -Hydroxy Esters to α -Keto Esters: GP 9. A dry three-necked round-bottom flask, equipped with a magnetic stirrer and a dropping funnel, was charged with anh. CH₂Cl₂ (7.5 ml/mmol) under N₂. After cooling to -78° (dry ice/acetone bath), oxalyl chloride (1.2 equiv.) and anh. DMSO (2 equiv.) were added dropwise, so that the temp. did not exceed -65° . The mixture was stirred at -78° for 10 min, treated with a soln. of the appropriate methyl ester (1 equiv.) in CH₂Cl₂ (1 ml/mmol), and stirred for additional 1.5 h at -78° . After addition of dry Et₃N (4 equiv.), the mixture was allowed to warm to r.t. over 0.5 h, whereupon H₂O (4 ml/mmol) was added. The phases were separated, and the aq. phase was extracted with CH₂Cl₂ (3×). The combined org. phases were washed with 1% HCl, 5% NaHCO₃, and brine, dried (MgSO₄), filtered, and evaporated. The crude keto esters are immedially used for the DAST reactions without prior purification.

Preparation of 6-Alkyl-2-(tert-butyl)tetrahydropyrimidin-4(IH)-ones: GP 10. At -20° , 1 equiv. of the N-Cbz-protected β-amino acid was dissolved in THF, and 1.2 equiv. of Et₃N and ethyl chloroformate were added. The resulting colorless suspension was cooled below -50° , and via a needle NH₃ gas was bubbled in for 1 h. After another 3 h, the solvent was removed by rotary evaporation. To the colorless solid, H₂O was added, and the resulting suspension was filtered and washed with H₂O and Et₂O. The isolated powder was dried for 12 h under h.v., then suspended in MeOH, and 10% Pd/C was added. After 12 h under H₂ (balloon), the Pd/C was filtered off (Celite), MeOH was evaporated, and the obtained β-amino acid amide was dissolved in CH₂Cl₂, 2 equiv. of pivalaldehyde was added, and the mixture was heated under reflux in an inverse Dean–Stark trap for 12 h. The solvent was removed, and the isolated crude product was purified by FC and/or recrystallization.

Cbz Protection of Tetrahydropyrimidin-4(1H)-ones: GP 11. N,O-Bis(trimethylsilyl)acetamide (1.5 equiv.) was added to a soln. of the corresponding tetrahydropyrimidin-4(1H)-one, $\mathbf{11}-\mathbf{13}$, in CH₂Cl₂ at r.t. After 1 h, the mixture was cooled to 0° and Cbz chloride (1.3 equiv.) was added. After 20 h at 0° , sat. NaHCO₃ was added, and the aq. layer was extracted with CH₂Cl₂ (3×). The combined org. layers were dried (MgSO₄), filtered, and evaporated. The crude products were purified by FC.

Peptide Coupling: GP 12. GP 12a. With HBTU. A soln. of the ammonium salt of the amino acid ester or peptide ester (1 equiv.) in CH_2Cl_2 was cooled to 0° , treated successively with the appropriate carboxylic acid (1 equiv.), NMM (3 equiv.), and HBTU (1.2 equiv.), and stirred for 12 h at r.t. The mixture was diluted with AcOH, and washed with aq. HCl (1M), sat. K_2CO_3 , and brine. The org. phase was dried (MgSO₄), filtered, evaporated, and the obtained crude product was purified by FC.

GP 12b. With EDC/HOBt. A soln. of the amino acid or peptide ester, or its TFA salt (1 equiv.) in CH_2Cl_2 (3 ml/mmol) was cooled to -10° , treated successively with a soln. of the appropriate acid (1 equiv.) in CH_2Cl_2 or THF (3 ml/mmol), NMM (3-5 equiv.), HOBt (1.2 equiv.) and EDC·HCl (1.2 equiv.), and stirred at -10° for 12 h. The mixture was diluted with AcOEt, washed with 1M HCl (3×),

sat. K_2CO_3 (3×), and brine. The org. layer was dried (MgSO₄), filtered, and evaporated, and the crude product was purified by FC.

GP 12c. With HATU. A soln. of the amino acid or peptide ester, or its TFA salt (1 equiv.) in CH_2Cl_2 or DMF (6 ml/mmol) was cooled to 0° , treated successively with the appropriate acid (1 equiv.), NMM (3–5 equiv.), and HATU (1.2 equiv.), and stirred at r.t. for 12 h. The mixture was diluted with AcOEt, washed with 1M HCl (3×), sat. K_2CO_3 (3×), and brine. The org. layer was dried (MgSO₄), filtered, and evaporated, and the crude product was purified by FC.

GP 12d. With HATU, Forming an Insoluble Peptide. The peptide-coupling reaction was performed according to GP 12c, but during the reaction the formed peptide precipitated. The mixture was evaporated, and the residue was stirred in AcOEt for 10 min. The resulting suspension was centrifuged, and the solid was stirred successively in AcOEt ($2\times$) and MeOH/H₂O 1:1 ($3\times$) for 10 min each. After the final centrifugation, the product was dried under h.v. for 12 h.

Cyclization of Oligopeptides: GP 13. A soln. of the unprotected peptide in DMF (0.4M) was treated at r.t. with pentafluorophenol (1 equiv.) and EDCI (1 equiv.). After 16 h, the mixture was evaporated, and the residue was dissolved in CHCl3. The resulting soln. was washed successively with 1M aq. HCl and brine. The org. phase was dried (MgSO4) and evaporated. The residue was dissolved in CH2Cl2 (0.5M) and an equal volume of TFA was added at 0° . The mixture was stirred for 1 h at 0° and for 1 h at r.t. The solvent was evaporated, and the residue was dissolved in toluene and evaporated twice. The obtained residue was dissolved in MeCN (0.025M) and slowly added (over 4 h) to a soln. of Hünig's base (EtNⁱPr2) in MeCN (3.3 mm) at 70° (bath temp.) with a syringe pump. The resulting precipitate was filtered, washed, and dried under h.v.

Boc Deprotection: GP 14. GP 14a. A soln. of the N-Boc-protected compound in CH_2Cl_2 (3 ml/mmol) was cooled to 0° (ice bath), treated with TFA (3 ml/mmol), and stirred at 0° for 1.5 h. After concentration under reduced pressure, the TFA salt was dried under h.v. for 2 h and used without further purification.

GP 14b. After Boc deprotection according to GP 14a, the TFA salt was dissolved in AcOEt, washed with sat. K_2CO_3 (2×), dried (MgSO₄), filtered, and evaporated. The resulting amine was used without further purification.

3. Preparation of the Fluorinated β -Amino Acid Derivatives 1–3, 7, and 8. 3.1. By Enolate Fluorination (Scheme 2, b). Methyl (2S,3S)-3-{[(tert-Butoxy)carbonyl]amino]-2-fluorobutanoate (1abb)}. Fluorination of Boc-hAla-methyl ester (1.87 g, 8.63 mmol; prepared from Boc-Ala [28]) was performed according to $GP\ 1$. Purification by FC (CH₂Cl₂/AcOEt 98:2) afforded 1abb (1.38 g, 68%). Pale yellow oil, which crystallized by scratching. Recrystallization gave colorless needles. M.p. 66–68° (hexane/Et₂O). $[\alpha]_D^{28} = -1.9$ (c = 1.0, MeOH). IR: 3369m, 3252m, 2983m, 2944m, 1758m, 1686m, 1338m, 1275m, 1247m, 1225m, 1164m, 1135m, 1112m, 1061m, 1008m, 980m, 938m, 921m, 877m, 849m, 783m, 750m, 696m, 675m. ¹H-NMR (400 MHz, CDCl₃): 1.16 (dd, d) = 7.0, 0.7, Me); 1.46 (d), 3 Me); 3.82 (d), Me); 4.12–4.40 (d), NCH); 4.79 (br. d), NH); 5.05 (dd), d) = 49.4, 2.2, CFH). ¹³C-NMR (100 MHz, CDCl₃): 14.2 (d), d) = 5.2, Me); 22.3 (3 Me); 47.8 (d), d) = 19.7, NCH); 52.5 (Me); 80.0 (CMe₃); 90.2 (d), d) = 188.2, FCH); 154.9 (CO); 168.2 (d), d) = 24.0, FCCO). ¹⁹F-NMR (280 MHz, CDCl₃): 40.6 (dd), d) = 49.0, 26.9, 1 F). Anal. calc. for C₁₀H₁₈FNO₄ (235.25): C 51.06, H 7.71, N 5.95; found: C 51.04, H 7.54, N 6.15.

Methyl (2S,3S)-3-{[(Benzyloxy)carbonyl]amino}-2-fluorobutanoate (**1acb**). Fluorination of Cbz-hAla methyl ester (5.03 g, 20.0 mmol, prepared from Cbz-Ala [28]) was performed according to *GP 1*. Purification by FC (CH₂Cl₂/AcOEt 98:2) afforded **1acb** (3.83 g, 71%). Pale yellow oil, which crystallized by scratching. Recrystallization gave colorless needles. M.p. $52-53^{\circ}$ (hexane/Et₂O). [α]²⁸₂ = +3.7 (c = 1.0, MeOH). IR: 3330m, 3068w, 3038w, 2992w, 2961w, 2896w, 2852w, 1759s, 1682s, 1652m, 1527s,

1466m, 1454m, 1437m, 1390w, 1381w, 1334m, 1276s, 1258m, 1228s, 1140m, 1113s, 1068m, 1013s, 983m, 941m, 912m, 860w, 840m, 785m, 751s, 730m, 696s, 671m, 655m. ¹H-NMR (300 MHz, CDCl₃): 1.17 (d, J = 6.9, Me); 3.81 (s, Me); 4.12 – 4.40 (m, NCH); 4.92 – 5.17 (m, NH, FCH, PhC H_2); 7.27 – 7.38 (m, 5 arom. H). ¹³C-NMR (100 MHz, CDCl₃): 14.3 (d, J = 5.3, Me); 48.3 (d, J = 20.4, NCH); 52.6 (Me); 67.1 (PhC H_2); 90.0 (d, J = 188.5, FCH); 128.2 (CH); 128.3 (CH); 128.6 (CH); 136.2 (CH); 155.4 (CO); 168.0 (d, J = 23.7, FCCO). ¹⁹F-NMR (282 MHz, CDCl₃): 41.0 (dd, J = 49.0, 26.7, 1 F). Anal. calc. for $C_{13}H_{16}FNO_4$ (269.27): C 66.84, H 8.56, N 4.10; found: C 66.57, H 8.46, N 4.11.

3.2. By the Cyanohydrin Route (Scheme 1). 3.2.1. Monofluorinated Ala-Derived Compounds. Methyl (2R,3S)-3-(Dibenzylamino)-2-fluorobutanoate (epi-7a) and Methyl (2R,3R)-2-(Dibenzylamino)-3-fluorobutanoate (iso-epi-7a). The methyl ester epi-6a [4] (2.04 g, 6.51 mmol) was dissolved in CH₂Cl₂ (13 ml) and fluorinated with DAST (1.3 ml, 8.70 mmol) at 0° for 3 h according to GP 2. FC (pentane/Et₂O 9:1) yielded epi-7a (753 mg, 37%) and iso-epi-7a (690 mg, 34%).

Data of epi-**7a.** Light yellow oil. $R_{\rm f}$ (pentane/Et₂O 5:1) 0.41. $[\alpha]_{\rm D}^{\rm LL} = +27.5$ (c=1.0, CHCl₃). IR (CHCl₃): 3570w, 3064w, 3032m, 2954w, 2841w, 2810w, 1762s, 1602w, 1495m, 1453m, 1439m, 1382m, 1358m, 1298m, 1171s, 1137w, 1106m, 1074w, 1025s, 948w, 911w, 832w. ¹H-NMR (400 MHz, CDCl₃): 1.27 (dd, J=0.6, 7.0, Me); 3.29 (ddq, J=3.8, 7.0, 31.8, NCH); 3.33 (d, J=13.4, 2 PhCHH'); 3.63 (s, MeO); 3.90 (d, J=13.4, 2 PhCHH'); 4.84 (dd, J=3.8, 49.1, CHF); 7.19 –7.32 (m, 10 arom. H). ¹³C-NMR (100 MHz, CDCl₃): 8.0 (d, J=4.4), 51.9 (Me); 53.9 (d, J=18.4, CH); 55.0 (CH₂); 94.2 (d, J=189.2), 127.0, 128.1, 129.1 (CH); 139.6, 169.0 (d, J=25.6) (C). ¹⁹F-NMR (282 MHz, CDCl₃): -200.4 (dd, J=32.0, 49.1, CHF). HR-MALDI-MS: 338.2 (11, [M+Na]+), 316.2 (100, [M+H]+), 314.2 (5), 296.2 (4), 268.2 (5), 224.1 (9), 158.1 (8). Anal. calc. for C₁₉H₂₂FNO₂ (315.39): C 72.36, H 7.03, N 4.44; found: C 72.51, H 7.08, N 4.35.

Data of iso-epi-**7a**. Light yellow oil. $R_{\rm f}$ (pentane/Et₂O 5 : 1) 0.52. [α]_D^{1,1} = -104.1 (c = 1.0, CHCl₃). IR (CHCl₃): 3063w, 3008m, 2953m, 2845w, 1729s, 1602w, 1495m, 1454m, 1435w, 1383w, 1359w, 1277w, 1162s, 115w, 1074m, 1024m, 990w, 939w, 877w, 843w. ¹H-NMR (400 MHz, CDCl₃): 1.35 (dd, J = 6.3, 24.0, Me); 3.37 (dd, J = 5.7, 23.7, NCH); 3.77 (d, J = 14.0, 2 PhCHH'); 3.78 (s, MeO); 4.06 (d, J = 14.0, 2 PhCHH'); 5.12 (ddq, J = 5.8, 6.3, 47.9, CHF); 7.21 – 7.40 (m, 10 arom. H). ¹³C-NMR (100 MHz, CDCl₃): 18.3 (d, J = 22.4), 51.4 (Me); 55.6 (CH₂); 64.9 (d, J = 19.6), 89.7 (d, J = 173.2), 127.1, 128.3, 128.8 (CH); 139.5, 171.0 (d, J = 6.7) (C). ¹⁹F-NMR (282 MHz, CDCl₃): -181.8 (dquint., J = 23.5, 48.0, CHF). HR-MALDI-MS: 338.2 (13, [M + Na]⁺), 316.2 (100, [M + H]⁺), 314.2 (15), 296.2 (7), 238.2 (14), 224.1 (28), 158.1 (8). Anal. calc. for C₁₉H₂₂FNO₂ (315.39): C 72.36, H 7.03, N 4.44; found: C 72.43, H 7.12, N 4.45

3.2.2. *Monofluorinated Val-Derived Compounds. Methyl* (2S,3S)-3-(*Dibenzylamino*)-2-hydroxy-4-methylpentanoate (**6b**). The cyanohydrin **5b** [12] (12.74 g, 41.3 mmol) was treated with a sat. HCl/MeOH soln. (210 ml) according to GP 8. FC (AcOEt/hexane 1:9 \rightarrow 3:7) yielded **6b** (8.61 g, 61%). Light yellow oil. R_f (CH₂Cl₂) 0.19. [a] $_D^{\text{Li}}$ = -0.5 (c = 1.0, CHCl₃). IR (CHCl₃): 3532w, 3064w, 2955m, 2873w, 2802w, 1728s, 1602w, 1494m, 1453m, 1366w, 1272m, 1137m, 1089m, 1028w, 990w, 967w, 913w. ¹H-NMR (400 MHz, CDCl₃): 0.71 (d, J = 6.6, Me); 1.02 (d, J = 6.7, Me); 2.18 – 2.31 (m, Me₂CH); 2.74 (dd, J = 1.5, 10.2, NCH); 2.99 (d, J = 4.6, OH); 3.40 (d, J = 13.9, 2 PhCHH'); 3.72 (s, MeO); 3.95 (d, J = 13.9, 2 PhCHH'); 4.59 (dd, J = 1.5, 4.6, CHOH); 7.20 – 7.38 (m, 10 arom. H). ¹³C-NMR (100 MHz, CDCl₃): 19.8, 21.5 (Me); 26.3 (CH); 52.5 (Me); 54.8 (CH₂); 66.5, 67.4, 126.9, 128.1, 129.2 (CH); 139.9, 176.7 (C). HR-MALDI-MS: 364.2 (9, [M + Na] $^+$), 342.2 (100, [M + H] $^+$), 252.2 (23). Anal. calc. for C₂₁H₂₇NO₃ (341.44): C 73.87, H 7.97, N 4.10; found: C 74.08, H 7.81, N 4.30.

Methyl (2S,3S)-3-(*Dibenzylamino*)-2-fluoro-4-methylpentanoate (**7b**). Compound **6b** (3.84 g, 11.25 mmol) was dissolved in CH₂Cl₂ (22 ml) and fluorinated with DAST (2.2 ml, 16.9 mmol) at 0° for 3 h, according to *GP* 2. FC (pentane/Et₂O 10:1) yielded **7b** (2.78 g, 72%). Yellow oil. $R_{\rm f}$ (pentane/Et₂O 10:1) 0.35. $[\alpha]_{\rm D}^{\rm FL} = -15.4$ (c = 1.0, CHCl₃). IR (CHCl₃): 3066w, 3032m, 2956m, 2803w, 1758s, 1602w,

1494m, 1476w, 1453m, 1438m, 1366w, 1287s, 1136m, 1116m, 1084s, 1016m, 992w, 968w, 912w, 867w. ¹H-NMR (400 MHz, CDCl₃): 0.80 (d, J = 6.6, Me); 1.00 (d, J = 6.7, Me); 2.13 – 2.26 (m, Me₂CH); 2.89 (ddd, J = 1.0, 9.9, 29.2, NCH); 3.34 (d, J = 13.8, 2 PhCHH'); 3.75 (s, MeO); 3.96 (d, J = 13.8, 2 PhCHH'); 5.37 (d, J = 48.3, CHF); 7.22 – 7.38 (m, 10 arom. H). ¹³C-NMR (100 MHz, CDCl₃): 19.3, 21.3 (Me); 26.3 (d, J = 5.0, CH); 52.3 (Me); 54.6 (CH₂); 66.0 (d, J = 18.9), 86.3 (d, J = 195.3), 127.1, 128.2, 129.2 (CH); 139.2, 171.3 (d, J = 23.1) (C). ¹⁹F-NMR (282 MHz, CDCl₃): -201.9 (dd, J = 28.8, 48.0, CHF). HR-MALDI-MS: 366.2 (1, [M + Na] $^+$), 344.2 (18, [M + H] $^+$), 300.1 (6), 252.2 (27). Anal. calc. for C₂₁H₂₆FNO₂ (343.44): C 73.44, H 7.63, N 4.08; found: C 73.30, H 7.47, N 4.01.

(2S,3S)-3-{[(tert-Butoxy)carbonyl]amino]-2-fluoro-4-methylpentanoic Acid (**1bba**). Compound **7b** (2.77 g, 8.06 mmol) was hydrolyzed with LiOH · H₂O (1.02 g, 24.2 mmol) in EtOH/H₂O (30 ml, 2 :1) according to GP 3, the carboxylic acid was dissolved in MeOH (80 ml) and hydrogenolyzed according to GP 4, and Boc-protected with Boc₂O (2.0 g, 9.17 mmol) and Et₃N (3.2 ml, 22.9 mmol) in MeOH (28 ml) according to GP 5. FC (CH₂Cl₂/MeOH/AcOH 100 :3 :0.1 → 100 :5 :0.2) yielded the carboxylic acid **1bba** (1.62 g, 80% over 3 steps). Colorless solid. M.p. 133 − 134°. R_f (CH₂Cl₂/MeOH/AcOH 100 :5 :1) 0.25. [α] $_{\rm B}^{\rm L}$ = −5.8 (c = 1.0, CHCl₃). IR (CHCl₃): 3447m, 3011m, 2980m, 2933m, 1749m, 1712m, 1503m, 1456m, 1393m, 1369m, 1161m, 1128m, 1098m, 1041m, 999m, 868m. ¹H-NMR (400 MHz, CD₃OD): 0.94 (d, d = 6.8, Me); 0.96 (d, d = 6.8, Me); 1.44 (m, m-Bu); 1.93 − 2.02 (m, Me₂CH); 3.91 (m, d = 5.5, 21.5, NCH); 4.89 (d, d = 5.1, 48.7, CHF). ¹³C-NMR (100 MHz, CD₃OD): 18.2, 20.5, 28.8 (Me); 30.0 (d, d = 2.6), 57.9 (d, d = 21.2) (CH); 80.4 (C); 91.2 (d, d = 186.9) (CH); 158.3, 172.1 (d, d = 23.1) (C). ¹⁹F-NMR (282 MHz, CD₃OD): −199.7 (dd, d = 21.3, 49.1, CHF). HR-ESI-MS: 565.2 (23), 543.2 (90, [2d + 2 Na]⁺), 521.3 (59, [2d + Na]⁺), 294.1 (23), 272.1 (100, [d + Na]⁺), 216.1 (14). Anal. calc. for C₁₁H₂₀FNO₄ (249.28): C 53.00, H 8.09, N 5.62; found: C 53.04, H 7.86, N 5.44.

Methyl (2S/R,3S)-3-(*Dibenzylamino*)-2-hydroxy-4-methylpentanoate (**6b**/epi-**6b**). A soln. of freshly prepared **4b** [11] (16.2 g, 57.5 mmol) in hexane/H₂O (115 ml, 3:1) was treated with acetone cyanohydrin (7.85 ml, 86.3 mmol), KCN (112.2 mg, 1.7 mmol), and Bu₄NI (148.7 mg, 0.4 mmol) according to *GP 7*. The crude epimeric mixture of cyanohydrins was treated with a sat. HCl/MeOH soln. (290 ml) according to *GP 8*. FC (hexane/AcOEt 98:2 → 7:3) yielded **6b**/epi-**6b** (15.5 g, 79% over 2 steps). Yellow oil. $R_{\rm f}$ (hexane/AcOEt 7:3) 0.46. The mixture could be separated by a further FC (hexane/AcOEt 95:5) to yielding pure **6b** and *epi-***6b**. The $^{\rm 1}$ H- and $^{\rm 13}$ C-NMR data for **6b** ((2*R*,3*S*)) were in accordance with those described above.

Data of epi-**6b** ((2S,3S)). Light yellow oil. $R_{\rm f}$ (pentane/Et₂O 3:2) 0.42. $[\alpha]_{\rm D}^{\rm pt}=-15.6$ (c=1.0, CHCl₃). IR (CHCl₃): 3600w, 3527w, 3067w, 3008m, 2961m, 2872w, 1949w, 1887w, 1815w, 1731s, 1494w, 1453m, 1389w, 1269s, 1146m, 1076m. $^{\rm 1}$ H-NMR (300 MHz, CDCl₃): 1.06 (d, J=6.5, Me); 2.35-2.47 (m, Me₂CH); 2.82 (dd, J=3.3, 8.6, NCH); 3.22 (br. s, OH); 3.44 (s, MeO); 3.75 (d, J=13.4, 2 PhCHH'); 3.99 (d, J=13.7, d PhCHH'); d (d); d (d), d (d), d), d (d), d), d0, d1, d3, d3, d4, d3, d4, d5, d5, d4, d5, d6, d7, d8, d9, d9,

321.2 (26), 291.2 (19), 266.2 (70), 252.2 (49). Anal. calc. for $C_{21}H_{27}NO_3$ (341.44): C 73.87, H 7.97, N 4.10; found: C 73.85, H 7.84, N 4.20.

Methyl (2R,3S)-3-(*Dibenzylamino*)-2-fluoro-4-methylpentanoate (epi-**7b**). Compound epi-**6b** (911 mg, 2.67 mmol) was dissolved in CH₂Cl₂ (6 ml) and fluorinated with DAST (525 μl, 4.00 mmol) at 0° for 3 h, according to *GP* 2. FC (pentane/Et₂O 99:1 → 97:3) yielded epi-**7b** (551 mg, 60%). Yellow oil. R_f (pentane/Et₂O 10:1) 0.21. [α]_D^{TL} = −22.8 (c = 1.0, CHCl₃). IR (CHCl₃): 3065w, 3032m, 2955s, 2851w, 1760s, 1602w, 1494m, 1476w, 1453s, 1390w, 1359m, 1294m, 1144s, 1116m, 1073s, 1016m, 979w, 949w, 911w, 835w. ¹H-NMR (500 MHz, CDCl₃): 1.02 (d, J = 6.6, Me); 1.12 (d, J = 6.8, Me); 2.29 − 2.37 (m, Me₂CH); 2.88 (ddd, J = 2.6, 9.0, 34.0, NCH); 3.63 (s, MeO); 3.81 (d, J = 13.6, 2 PhCHH'); 5.15 (dd, J = 2.6, 47.9, CHF); 7.19 − 7.30 (m, 10 arom. H). ¹³C-NMR (125 MHz, CDCl₃): 21.2, 22.1 (Me); 27.8 (d, J = 1.8, CH); 52.0 (Me); 55.6 (CH₂); 64.5 (d, J = 18.0), 91.1 (d, J = 189.6), 126.9, 128.1, 129.4 (CH); 140.0, 170.2 (d, J = 25.3) (C). ¹⁹F-NMR (282 MHz, CDCl₃): −202.1 (dd, J = 34.2, 48.0, CHF). HR-MALDI-MS: 366.2 (5, [M + Na]⁺), 344.2 (71, [M + H]⁺), 324.2 (12), 252.2 (100), 126.1 (7). Anal. calc. for C₂₁H₂₆FNO₂ (343.44): C 73.44, H 7.63, N 4.08; found: C 73.49, H 7.59, N 4.20.

(2R,3S)-3-{[(tert-Butoxy)carbonyl]amino}-2-fluoro-4-methylpentanoic Acid (**2bba**). Compound epi-**7b** (472 mg, 1.37 mmol) was hydrolyzed with LiOH·H₂O (173 mg, 4.12 mmol) in EtOH/H₂O (7.5 ml, 2:1) according to GP 3, the carboxylic acid was dissolved in MeOH (15 ml), hydrogenolyzed according to GP 4, and Boc-protected with Boc₂O (354 mg, 1.64 mmol) and Et₃N (570 µl, 4.1 mmol) in MeOH (5 ml) according to GP 5. FC (hexane/AcOEt 2:1+2% AcOH) yielded **2bba** (293 mg, 85% over 3 steps). Light yellow solid. M.p. 70-73°. R_f (CH₂Cl₂/MeOH/AcOH 100:5:1) 0.27. [α] $_D^{\text{pi}}$ -= -35.8 (c= 1.1, CHCl₃). IR (CHCl₃): 3440m, 3026w, 2979m, 2923w, 1754w, 1713s, 1505s, 1456w, 1393w, 1369m, 1310w, 1161s, 1077w, 1045w, 904w, 861w. ¹H-NMR (300 MHz, CD₃OD): 0.97 (d, J = 6.8, Me); 1.03 (d, J = 6.5, Me); 1.42 (s, t-Bu); 1.83 – 1.95 (m, Me₂CH); 3.80 (ddd, J = 1.9, 8.7, 30.2, NCH); 5.13 (dd, J = 1.9, 48.2, CHF). ¹³C-NMR (75 MHz, CD₃OD): 18.4, 18.8, 27.5 (Me); 30.1, 57.8 (d, J = 18.9) (CH); 79.0 (C); 88.5 (d, J = 185.6, CH); 156.9, 177.9 (C). ¹⁹F-NMR (282 MHz, CD₃OD): -203.2 (dd, J = 29.9, 48.0, CHF). HR-ESI-MS: 565.2 (27), 543.2 (14, [2M + 2 Na]⁺), 294.1 (100), 272.1 (33, [M + Na]⁺). Anal. calc. for C₁₁H₂₀FNO₄ (249.28): C 53.00, H 8.09, N 5.62; found: C 52.96, H 7.96, N 5.43.

3.2.3. Monofluorinated Leu-Derived Compounds. Methyl (2S,3S)-3-(Dibenzylamino)-2-hydroxy-5-methylhexanoate (**6c**). The cyanohydrin $\mathbf{5c}$ [12] (4.43 g, 13.8 mmol) was treated with a sat. HCl/MeOH soln. (70 ml) according to GP 8. FC (AcOEt/hexane $1:9 \rightarrow 3:7$) yielded $\mathbf{6c}$ (4.20 g, 86%). Light yellow oil. $R_{\rm f}$ (pentane/Et₂O 4:1) 0.23. [a] $_{\rm in}^{\rm ft}$ = +10.5 (c = 1.0, CHCl₃). IR (CHCl₃): 3528w, 3032w, 2956m, 2869w, 2810w, 1729s, 1602w, 1494m, 1453m, 1366w, 1268m, 1137w, 1089m, 1071m, 966w, 905w. $^{\rm th}$ -NMR (400 MHz, CDCl₃): 0.52 (d, J = 6.5, Me); 0.84 (d, J = 6.7, Me); 0.92 (ddd, J = 4.9, 8.4, 13.7, 1 H, CH₂CH); 1.67 (ddd, J = 5.0, 8.8, 14.0, 1 H, CH₂CH); 1.74 – 1.80 (m, Me₂CH); 3.04 – 3.08 (m, NCH); 3.05 (d, J = 6.8, OH); 3.50 (d, J = 13.8, 2 PhCHH'); 3.71 (s, MeO); 3.86 (d, J = 13.8, 2 PhCHH'); 4.61 (dd, J = 2.4, 6.7,

CHOH); 7.20-7.35 (m, 10 arom. H). 13 C-NMR (100 MHz, CDCl₃): 21.7, 23.4 (Me); 24.2 (CH); 35.0 (CH₂); 52.4 (Me); 54.5 (CH₂); 57.9, 69.3, 127.0, 128.2, 129.1 (CH); 140.0, 175.6 (C). HR-MALDI-MS: 378.2 (6, $[M+Na]^+$), 356.2 (100, $[M+H]^+$), 266.2 (50). Anal. calc. for $C_{22}H_{29}NO_3$ (355.48): C 74.33, H 8.22, N 3.94; found: C 74.15, H 8.08, N 4.02.

Methyl (2S,3S)-3-(*Dibenzylamino*)-2-fluoro-5-methylhexanoate (**7c**). Compound **6c** (325 mg, 0.91 mmol) was dissolved in CH₂Cl₂ (2 ml) and fluorinated with DAST (180 µl, 1.37 mmol) at 0° for 3 h, according to *GP* 2. FC (pentane/Et₂O 99:1 → 97:3) yielded **7c** (277 mg, 85%). Light yellow oil. $R_{\rm f}$ (pentane/Et₂O 10:1) 0.24. [α]_b: = −6.6 (c = 1.0, CHCl₃). IR (CHCl₃): 3065w, 3032w, 2956m, 2869w, 2806w, 1756s, 1603w, 1495m, 1454m, 1439m, 1367m, 1285s, 1162w, 1138w, 1086w, 1070m, 1017w, 967w, 907w, 868w. ¹H-NMR (400 MHz, CDCl₃): 0.44 (d, d = 6.4, Me); 0.84 (d, d = 6.6, Me); 0.96−1.03 (m, 1 H, CH₂CH); 1.74−1.84 (m, Me₂CH, 1 H of CH₂CH); 3.16 (dddd, d = 1.8, 3.8, 9.7, 31.3, NCH); 3.44 (d, d = 13.8, 2 PhCHH'), 3.74 (s, MeO); 3.94 (d, d = 13.8, 2 PhCHH'); 5.42 (dd, d = 1.8, 50.3, CHF); 7.21−7.35 (m, 10 arom. H). ¹³C-NMR (100 MHz, CDCl₃): 21.1, 23.6 (Me); 24.0, 35.2 (d, d = 4.0) (CH); 52.3 (Me); 54.2, 54.3 (CH₂); 57.3 (d, d = 19.1), 88.0 (d, d = 191.7), 127.1, 128.2, 129.1 (CH); 139.5, 170.2 (d, d = 24.0) (C). ¹⁹F-NMR (282 MHz, CDCl₃): −201.6 (ddd, d = 30.9, 50.2, CHF). HR-MALDI-MS: 397.0 (7, [d + K]⁺), 380.2 (12, [d + Na]⁺), 358.2 (100, [d + H]⁺), 354.1 (26), 338.2 (17), 280.2 (45), 266.2 (60). Anal. calc. for C₂₂H₂₈FNO₂ (357.47): C 73.92, H 7.89, N 3.92; found: C 73.80, H 7.73, N 3.96.

Benzyl (2S,3S)-3-{[(tert-Butoxy)carbonyl]amino}-2-fluoro-5-methylhexanoate (1cbc). Compound **7c** (4.13 g, 11.55 mmol) was hydrolyzed with LiOH \cdot H₂O (1.45 g, 34.65 mmol) in EtOH/H₂O (45 ml, 2:1) according to GP 3; the obtained carboxylic acid was dissolved in MeOH (115 ml) and hydrogenolyzed according to GP 4, and Boc-protected with Boc₂O (3.02 g, 13.86 mmol) and Et₃N (4.87 ml, 34.65 mmol) in MeOH (35 ml) according to GP 5. The resulting acid 1cba was dissolved in DMF (35 ml) and treated with Cs₂CO₃ (3.78 g, 11.6 mmol) and BnBr (1.66 ml, 13.9 mmol) according to GP 6. FC (pentane/Et₂O 7:1) yielded **1cbc** (3.43 g, 84% over 4 steps). Colorless oil. R_f (pentane/AcOEt 9:1) 0.44. $[\alpha]_D^{\text{t.}} = -18.2$ $(c = 1.0, CHCl_2)$. IR $(CHCl_3)$: 3446w, 3036w, 2964m, 2872w, 1759m, 1708s, 1503s, 1456w, 1390w, 1369m, 1164s, 1128w, 1103w, 1046w, 908w. 1 H-NMR (400 MHz, CDCl₃): 0.75 (d, J = 6.5, Me); 0.83 (d, J = 6.7, Me); $0.96 (ddd, J = 3.1, 10.0, 13.3, 1 \text{ H}, CH_2CH)$; $1.39 - 1.46 (m, 1 \text{ H}, CH_2CH)$; 1.44 (s, t-Bu); 1.54 - 1.59 (m, Me_2CH) ; 4.09 – 4.22 (m, NCH); 4.59 (d, J = 9.1, BocNH); 5.06 (dd, J = 2.6, 49.2, CHF); 5.15 (d, J = 4.0, CHF); 5.16 (d, J = 4.0, CHF); 5.16 (d, J = 4.0, CHF); 5.17 (d, J = 4.0, CHF); 5.17 (d, J = 4.0, CHF); 5.18 (d, J = 4.0, CHF); 5.19 (d, J = 4.0, CHF); 5.19 (d, J = 4.0, CHF); 6.18 (d, J = 4.0, CHF)11.9, 1 H, PhC H_2); 5.32 (d, J = 11.9, 1 H, PhC H_2); 7.33 – 7.39 (m, 5 arom. H). ¹³C-NMR (100 MHz, $CDCl_3$): 21.1, 23.4 (Me); 24.4 (CH); 28.3 (Me); 37.4 (d, J = 3.3, CH_2); 50.3 (d, J = 19.7, CH); 67.3 (CH_2); 79.9 (C); 90.6 (d, J = 187.5), 128.7, 128.8, 128.9 (CH); 134.9, 155.2, 167.6 (d, J = 24.2) (C). ¹⁹F-NMR $(282 \text{ MHz}, \text{CDCl}_3): -199.0 (dd, J = 26.7, 48.0, \text{CHF}). \text{ HR-MALDI-MS: } 392.2 (7, [M+K]^+), 376.2 (100, 100)$ [M+Na]+), 344.2 (12), 320.1 (60), 300.1 (18), 254.2 (48). Anal. calc. for C₁₉H₂₈FNO₄ (353.43): C 64.57, H 7.98, N 3.96; found: C 64.51, H 7.82, N 3.91.

Methyl (2S/R,3S)-3-(*Dibenzylamino*)-2-hydroxy-5-methylhexanoate (**6c**/epi-**6c**). A soln. of freshly prepared **4c** [11] (5.6 g, 19.1 mmol) in hexane/H₂O (40 ml, 3:1) was treated with acetone cyanohydrin (2.6 ml, 28.7 mmol), KCN (40.0 mg, 0.6 mmol), and Bu₄NI (47.0 mg, 0.2 mmol) according to *GP 7*. The crude epimeric mixture of cyanohydrins was treated with a sat. HCl/MeOH soln. (100 ml) according to *GP 8*. FC (pentane/Et₂O 4:1) yielded **6c**/epi-**6c** (7.12 g, 78% over 2 steps). Yellow oil. $R_{\rm f}$ (pentane/Et₂O 4:1) 0.23. Both epimers could be separated by a further FC (hexane/AcOEt 95:5) yielding pure **6c** and epi-**6c**. The ¹H- and ¹³C-NMR data for **6c** ((2*R*,3*S*)) were in accordance with those described above.

Data of epi-**6c** ((2S,3S)). Light yellow oil. $R_{\rm f}$ (pentane/Et₂O 3:2) 0.43. [α]₅L = +43.7 (c = 1.0, CHCl₃). IR (CHCl₃): 3520w, 3008w, 2957s, 1731s, 1495m, 1454m, 1366w, 1269m, 1155m, 1092m, 1028w, 973w. ¹H-NMR (300 MHz, CDCl₃): 0.94 (d, J = 6.5, Me); 1.00 (d, J = 6.2, Me); 1.48 – 1.58 (m, 1 H, CH₂CH); 1.60 – 1.69 (m, 1 H, CH₂CH); 1.77 – 1.86 (m, Me₂CH); 3.00 – 3.08 (m, NCH); 3.36 (d, J = 13.7, 2 PhCHH'); 3.48 (s, MeO); 3.95 (d, J = 13.4, 2 PhCHH'); 4.18 (d, J = 4.1, CHOH); 7.19 – 7.39 (m, 10 arom. H). ¹³C-NMR (75 MHz, CDCl₃): 22.1, 24.1 (Me); 25.5 (CH); 32.8 (CH₂); 52.4 (Me); 55.6 (CH₂); 57.2, 73.2, 126.8, 128.1, 129.1 (CH); 139.9, 174.9 (C). HR-MALDI-MS: 378.2 (9, [M + Na]⁺), 356.2 (100, [M + H]⁺), 266.2 (60), 178.1 (9), 176.1 (8).

Methyl (2R,3S)-3-(*Dibenzylamino*)-2-fluoro-5-methylhexanoate (epi-**7c**). Compound epi-**6c** (2.42 g, 6.80 mmol) was dissolved in CH₂Cl₂ (14 ml) and fluorinated with DAST (1.34 ml, 10.21 mmol) at 0° for 3 h, according to *GP* 2. FC (pentane/Et₂O 99:1 → 96:4) yielded epi-**7c** (1.09 g, 45%). Light yellow solid. M.p. 62−63°. $R_{\rm f}$ (pentane/Et₂O 10:1) 0.31. [a]_D^{1,1} = +31.7 (c=1.1, CHCl₃). IR (CHCl₃): 3064w, 3032w, 2957s, 2870w, 2810w, 1762s, 1603w, 1495m, 1454m, 1439w, 1361w, 1304m, 1158m, 1076m, 1029w, 1001w, 972w, 938w, 911w, 887w, 831w. ¹H-NMR (400 MHz, CDCl₃): 0.89 (d, J = 6.4, Me); 0.92 (d, J = 6.4, Me); 1.50−1.76 (m, Me₂CH₂CH); 3.15 (tdd, J = 4.0, 10.2, 31.4, NCH); 3.40 (d, J = 13.4, 2 PhCHH'); 3.61 (s, MeO); 3.87 (d, J = 10.3, 2 PhCHH'); 4.97 (dd, J = 3.6, 49.0, CHF); 7.19−7.31 (m, 10 arom. H). ¹³C-NMR (100 MHz, CDCl₃): 22.0, 23.7 (Me); 25.0 (CH); 31.9 (d, J = 3.3, CH₂); 51.9 (Me); 55.0, 55.1 (CH₂); 56.5 (d, J = 18.1), 91.9 (d, J = 188.8), 126.9, 128.1, 129.3 (CH); 139.7, 169.4 (d, J = 25.6) (C). ¹°F-NMR (282 MHz, CDCl₃): −202.6 (dd, J = 31.0, 48.0, CHF). HR-MALDI-MS: 358.2 (22, [M+H]⁺), 338.2 (62), 278.2 (19), 266.2 (100). Anal. calc. for C₂₂H₂₈FNO₂ (357.47): C 73.92, H 7.89, N 3.92; found: C 73.87, H 8.13, N 4.08.

Benzyl (2R,3S)-3-{[[(tert-Butoxy)carbonyl]amino]-2-fluoro-5-methylhexanoate (2cbc). Compound epi-7c (940 mg, 2.63 mmol) was hydrolyzed with LiOH·H₂O (331 mg, 7.89 mmol) in EtOH/H₂O 2:1 (10 ml) according to GP 3, the carboxylic acid was dissolved in MeOH (26 ml) and hydrogenolyzed according to GP 4, and Boc-protected with Boc₂O (707 mg, 3.24 mmol) and Et₃N (1.13 ml, 8.10 mmol) in MeOH (15 ml) according to GP 5. The resulting acid 2cba was dissolved in DMF (8 ml), and treated with Cs₂CO₃ (968 mg, 3.0 mmol) and BnBr (385 μl, 3.24 mmol) according to GP 6. FC (pentane/Et₂O 9:1) yielded 2cbc (861 mg, 92% over 4 steps). Colorless solid. M.p. 80–82°. R_f (pentane/AcOEt 9:1) 0.42. [α] $_D^{\text{in}}$ = -37.1 (c = 1.0, CHCl₃). IR (CHCl₃): 3439m, 3009w, 2962m, 2923w, 2872w, 1759s, 1709s, 1503s, 1456w, 1392w, 1368m, 1331w, 1164s, 1135w, 1083m, 1052w, 949w, 913w, 867w, 850w. ¹H-NMR (400 MHz, CDCl₃): 0.94 (d, J = 6.5, 2 Me); 1.36–1.50 (m, CH₂CH); 1.42 (s, t-Bu); 1.62–1.72 (m, Me₂CH); 4.20–4.33 (m, NCH); 4.66 (d, J = 10.0, BocNH); 4.89 (dd, J = 1.9, 47.9, CHF); 5.11 (d, J = 12.0, 1 H, PhCH₂); 5.27 (d, J = 12.0, 1 H, PhCH₂); 7.32–7.42 (m, 5 arom. H). ¹³C-NMR (100 MHz, CDCl₃): 22.0, 22.9 (Me); 24.7 (CH); 28.3 (Me); 40.3 (CH₂); 50.3 (d, J = 20.4, CH); 67.5 (CH₂); 79.7 (C); 89.7 (d, J = 187.4), 128.6, 128.7, 128.7 (CH); 135.0, 155.2, 168.1 (d, J = 25.2) (C). ¹⁹F-NMR (282 MHz, CDCl₃): -204.1 (dd, J = 26.7, 48.0,

CHF). HR-MALDI-MS: 376.2 (51, $[M+Na]^+$), 320.1 (34). Anal. calc. for $C_{19}H_{28}FNO_4$ (353.43): C 64.57, H 7.98, N 3.96, F 5.38; found: C 64.77, H 7.81, N 3.90, F 5.37.

3.2.4. Difluoro-Substituted Val- and Leu-Derived Compounds. Methyl (3S)-3-(Dibenzylamino)-2,2-difluoro-4-methylpentanoate (**8b**). The mixture **6b**/epi-**6b** (5.1 g, 14.9 mmol) was oxidized with oxalyl chloride (1.5 ml, 17.9 mmol) and DMSO (2.1 ml, 29.8 mmol) according to GP 9. The resulting crude keto ester was dissolved in CH_2Cl_2 (26 ml) and fluorinated with DAST (5.5 ml, 44.7 mmol) at r.t. for 6 h, according to GP 2. FC (pentane/Et₂O 95:5) yielded **8b** (3.4 g, 64% over 2 steps). Light yellow oil. R_f (pentane/Et₂O 95:5) 0.31. [α] $_D^{\text{pl}} = -2.1$ (c = 1.0, $CHCl_3$). IR ($CHCl_3$): 3005m, 2954m, 2841w, 1959w, 1769s, 1708s, 1600w, 1492w, 1451m, 1440w, 1415w, 1359s, 1303m, 1281w, 1164w, 1118s, 1087m, 1067s, 1041m, 979w, 913w, 841w. 11 H-NMR (500 MHz, $CDCl_3$): 0.96 (d, d) = 6.7, d); 1.06 (d, d) = 6.9, d); 1.3-2.19 (d), 1.3-3.25 (d), 1.3-3.25

$$Bn_2N$$
 F F OMe Boc N F F OMe Boc N F F OMe Boc N F F OMe OMe

(3S)-3-{[(tert-Butoxy)carbonyl]amino}-2,2-difluoro-4-methylpentanoic Acid (3bba). Compound 8b (847 mg, 3.0 mmol) was hydrolyzed with LiOH·H₂O (379 mg, 9.0 mmol) in EtOH/H₂O 2:1 (12 ml) according to GP 3. The resulting acid 3bba (790 mg, 98%) was used without further purification. Colorless solid. M.p. $115-116^{\circ}$. [a] $_{\rm D}^{\rm DL}=-49.4$ (c=1.0, CHCl $_{3}$). IR (CHCl $_{3}$): 3446m, 3026w, 2982m, 2933w, 1760m, 1717s, 1506s, 1394w, 1369m, 1310w, 1160s, 1092w, 1041w, 1005w, 873m. 1 H-NMR (300 MHz, CD $_{3}$ OD): 0.94 (d, J=6.8, Me); 0.98 (d, J=6.8, Me); 1.44 (s, t-Bu); 2.01-2.09 (m, Me₂CH); 4.11 (ddd, J=5.0, 13.1, 17.4, BocNHCH). 13 C-NMR (75 MHz, CD $_{3}$ OD): 17.6, 20.6, 28.4 (Me); 28.7, 57.8 (t, J=22.0) (CH); 80.2, 116.5 (t, J=253.9), 157.8, 166.0 (t, J=31.7) (C). 19 F-NMR (282 MHz, CD $_{3}$ OD): -111.1 (dd, J=12.8, 254.0, 1 F, CF $_{2}$); -112.6 (dd, J=17.1, 255.0, 1 F, CF $_{2}$). HR-ESI-MS: 613.3 (7, [2M+2 K] $^{+}$), 579.2 (12, [2M+2 Na] $^{+}$), 557.2 (100, [2M+Na] $^{+}$), 370.1 (15), 290.1 (64, [M+Na] $^{+}$), 234.1 (7), 212.1 (13).

Methyl (3S)-3-{[(tert-Butoxy)carbonyl]amino}-2,2-difluoro-4-methylpentanoate (**3bbb**). The hydrogenolysis of **8b** (2.0 g, 5.5 mmol) was performed in presence of Boc₂O (1.8 g, 8.2 mmol) in MeOH (90 ml) according to *GP* 4. FC (pentane/Et₂O 95:5) yielded **3bbb** (1.3 g, 83%). Colorless solid. $R_{\rm f}$ (pentane/Et₂O 95:5) 0.16. M.p. 48−49°. [a]_D¹ = −19.8 (c = 1.0, CHCl₃). IR (CHCl₃): 3446m, 2974m, 2882m, 1769m, 1713m, 1497m, 1446m, 1395m, 1369m, 1308m, 1159m, 1092m, 1056m, 1005m, 979m, 867m, 836m. ¹H-NMR (500 MHz, CDCl₃): 0.95 (d, d = 6.8, Me); 1.01 (d, d = 6.8, Me); 1.44 (g, d = 0.7) m, 1005 (g = 0.7) m, 111, 20.5 (Me); 2.11 (CH); 28.2, 53.4 (Me); 56.5 (dd, d = 21.7, 26.4, CH); 80.2, 115.2 (dd, d = 257.1), 155.4, 164.1 (dd, d = 31.2) (C). ¹⁹F-NMR (282 MHz, CDCl₃): −111.8 (dd, d = 8.5, 257.2, 1 F, CF₂); −116.8 (dd, d = 21.3, 257.2, 1 F, CF₂). HR-ESI-MS: 585.3 (39, [d + Na]⁺), 320.1 (9, [d + K]⁺), 304.1 (100, [d + Na]⁺). Anal. calc. for C₁₂H₂₁F₂NO₄ (281.30): C 51.24, H 7.52, N 4.98, F 13.51; found: C 51.41, H 7.46, N 5.14, F 13.51.

Methyl (3S)-3-(*Dibenzylamino*)-2,2-difluoro-5-methylhexanoate (8c). Mixture 6c/epi-6c (5.3 g, 15.0 mmol) was oxidized with oxalyl chloride (1.6 ml, 18.0 mmol) and DMSO (2.1 ml, 30.0 mmol) according to *GP* 9. The resulting crude keto ester was dissolved in CH₂Cl₂ (30 ml) and fluorinated with DAST (5.5 ml, 45.0 mmol) at r.t. for 6 h, according to *GP* 2. FC (pentane/CH₂Cl₂ 95:5) yielded 8c (3.7 g, 65% over 2 steps). Colorless solid. $R_{\rm f}$ (pentane/CH₂Cl₂ 95:5) 0.16. M.p. 60°. [a] $_{\rm ft}^{\rm ht}$ = +14.1 (c = 1.0, CHCl₃). IR (CHCl₃): 3087w, 3032w, 2958m, 2869m, 1767s, 1603w, 1496w, 1454m, 1440w, 1380w, 1313m, 1117s, 1062s, 1028w, 966w, 917w. $_{\rm ft}^{\rm ht}$ H-NMR (400 MHz, CDCl₃): 0.75 (d, J = 6.5, Me); 0.93 (d, J = 6.6,

Me); 1.45-1.63 (m, CH_2 CH); 1.79-1.88 (m, Me_2 CH); 3.31-3.42 (m, NCH); 3.56 (d, J=13.4, 2 PhCHH'); 3.68 (s, MeO); 3.82 (d, J=13.4, 2 PhCHH'); 7.20-7.32 (m, 10 arom. H). 13 C-NMR (100 MHz, CDCl₃): 22.6, 22.8 (Me); 24.9 (CH); 32.6 (d, J=2.5, CH₂); 52.9 (Me); 54.6 (CH₂); 57.0 (dd, J=20.4, 25.0, CH); 118.7 (t, J=258.0, C); 127.1, 128.2, 129.4 (CH); 139.2, 164.8 (dd, J=30.9, 34.0) (C). 19 F-NMR (282 MHz, CDCl₃): -103.7 (dd, J=11.7, 255.0, 1 F, CF₂); -114.2 (dd, J=23.5, 254.0, 1 F, CF₂). HR-MALDI-MS: 413.3 (10, $[M+K]^+$), 398.2 (22, $[M+Na]^+$), 376.2 (100, $[M+H]^+$), 284.2 (11), 266.2 (15), 181.1 (17). Anal. calc. for $C_{22}H_{27}F_2NO_2$ (375.46): C 70.38, H 7.25, N 3.73; found: C 70.43, H 7.40, N 3.72.

Methyl (*3*S)-*3*-{[(tert-Butoxy)carbonyl]amino}-2,2-difluoro-5-methylhexanoate (**3cbb**). The hydrogenolysis of **8c** (1.6 g, 4.2 mmol) was performed in presence of Boc₂O (1.3 g, 6.2 mmol) in MeOH (70 ml) according to *GP 4*. FC (pentane/Et₂O 19:1) yielded **3cbb** (1.3 g, 95%). Colorless solid. $R_{\rm f}$ (pentane/Et₂O 19:1) 0.17. M.p. 77°. [a]_L¹ = −39.6 (c = 1.0, CHCl₃). IR (CHCl₃): 3436w, 2964w, 2872w, 1769s, 1713s, 1503s, 1441w, 1390w, 1369w, 1308w, 1159s, 1072w, 1046w, 1021w, 959w, 877w, 846w, 826w. ¹H-NMR (500 MHz, CDCl₃): 0.93 (d, J = 6.5, Me); 0.96 (d, J = 6.7, Me); 1.42 (s, t-Bu); 1.36−1.46 (m, CH₂CH); 1.67−1.74 (m, Me₂CH); 3.86 (s, MeO); 4.25−4.35 (m, NCH); 4.51 (d, J = 10.0, BocNH). ¹³C-NMR (125 MHz, CDCl₃): 21.2, 23.5 (Me); 24.3 (CH); 28.2 (Me); 36.3 (CH₂); 51.2 (dd, J = 23.4, 27.4, CH); 53.4 (Me); 80.2, 114.7 (t, J = 254.3), 155.1, 163.9 (dd, J = 31.3, 33.5) (C). ¹⁹F-NMR (282 MHz, CDCl₃): −103.7 (dd, J = 11.7, 254.0, 1 F, CF₂); −114.2 (dd, J = 18.1, 254.0, 1 F, CF₂). HR-ESI-MS: 660.9 (4), 613.3 (100, [2M + Na]⁺), 318.1 (53, [M + Na]⁺), 306.6 (5), 214.8 (3), 209.5 (4), 156.2 (4). Anal. calc. for C₁₃H₂₃F₂NO₄ (295.33): C 52.87, H 7.85, N 4.74; found: C 52.86, H 7.65, N 4.79.

Benzyl (3S)-3-{[(tert-Butoxy)carbonyl]amino}-2,2-difluoro-5-methylhexanoate (**3cbc**). Compound **3cbb** (1.13 g, 4.02 mmol) was hydrolyzed with LiOH·H₂O (506 mg, 12.6 mmol) in EtOH/H₂O 2:1 (15 ml) according to GP 3. The resulting carboxylic acid **3cba** was dissolved in DMF (12 ml) and treated with Cs₂CO₃ (1.31 g, 4.02 mmol) and BnBr (570 µl, 4.82 mmol) according to GP 6. FC (pentane/Et₂O 19:1) yielded **3cbc** (1.40 g, 96% over 2 steps). Colorless solid. M.p. 47°. R_f (pentane/Et₂O 19:1) 0.20. [α] $_{0}^{+}$ = -18.6 (c = 1.0, CHCl₃). IR (CHCl₃): 3436m, 3026w, 2954m, 2861w, 1764s, 1713s, 1503s, 1456m, 1390w, 1369s, 1303w, 1267w, 1159s, 1072s, 1046m, 1026w, 954w, 908w, 877w, 846w. ¹H-NMR (500 MHz, CDCl₃): 0.87 (d, J = 6.5, Me); 0.91 (d, J = 6.7, Me); 1.31 – 1.37 (m, CH₂CH); 1.43 (s, t-Bu); 1.64 – 1.72 (m, Me₂CH); 4.23 – 4.39 (m, NCH); 4.51 (d, J = 10.3, BocNH); 5.19 (d, J = 12.0, 1 H, PhCH₂); 5.32 (d, J = 12.0, 1 H, PhCH₂); 7.34 – 7.47 (m, 5 arom. H). ¹³C-NMR (125 MHz, CDCl₃): 21.2, 23.4 (Me); 24.3 (CH); 28.2 (Me); 36.6 (CH₂); 51.2 (t, J = 26.3, CH); 68.5 (CH₂); 80.2, 114.7 (t, J = 255.9) (C); 128.7, 128.8 (CH); 134.2, 155.1, 163.3 (t, J = 31.8) (C). ¹⁹F-NMR (282 MHz, CDCl₃): -116.7 (dd, J = 11.7, 254.0, 1 F, CF₂); -118.8 (dd, J = 14.9, 252.9, 1 F, CF₂). HR-ESI-MS: 765.4 (18, [2M + Na]⁺), 410.2 (21, [M + K]⁺), 394.2 (100, [M + Na]⁺). Anal. calc. for C₁₉H₂₇F₂NO₄ (371.42): C 61.44, H 7.33, N 3.77, F 10.23; found: C 61.36, H 7.16, N 3.69, F 10.26.

4. Preparation of the Fluorinated Tetrahydropyrimidin-4(1H)-ones **11**–**13** (Fig. 2). (2S,5S,6S)-2-(tert-Butyl)-5-fluorotetrahydro-6-methylpyrimidin-4(1H)-ones **(11)**. According to $GP\ 10$, aminobutanoic acid **1aca** (1.82 g, 7.13 mmol) was converted to the corresponding Cbz-protected amino acid amide **1acd** (1.11 g, 61% yield), and subsequent hydrogenation (1.00 g, 3.93 mmol) gave the β -amino acid amide **1aad** (470 mg, quant.). Amide **1aad** (60.0 mg, 0.50 mmol) was treated with pivalaldehyde to afford crude **11** as colorless solid. Recrystallization of this crude product from hexane/AcOEt afforded **11** (81.1 mg, 86% yield). Colorless solid. A second recrystallization gave colorless prisms. M.p. 131–132° (hexane/AcOEt). [α] $_{20}^{16} = +21.0$ (c=1.0, CHCl $_{3}$). IR: 3318w, 3243m, 3107w, 2967m, 2954m, 2914m, 2871m, 1681s, 1660s, 1478s, 1454m, 1438m, 1417m, 1403m, 1381w, 1365m, 1330m, 1300m, 1284m, 1250m, 1207w, 1152m, 1134m, 1100m, 1056s, 1039s, 1017m, 965m, 937w, 904w, 874w, 805m, 763s, 686s, 635m. 1 H-NMR (400 MHz,

CDCl₃): 0.96 (s, 3 Me); 1.33 (d, J = 6.3, Me); 3.07 – 3.19 (m, NCH); 4.08 (d, J = 7.0, NCHN); 4.26 (dd, J = 48.3, 10.0, CFH); 6.17 (br. s, NH). ¹³C-NMR (100 MHz, CDCl₃): 18.6 (Me); 24.8 (3 Me); 34.2 (t-Bu); 51.7 (d, J = 21.9, NCH); 75.5 (NCHN); 88.9 (d, J = 189.9, FCH); 168.9 (d, J = 20.5, CFCO). ¹⁹F-NMR (376 MHz, CDCl₃): - 198.5 (dd, J = 48.4, 5.5, F). Anal. calc. for C₉H₁₇FN₂O (188.24): C 57.42, H 9.10, N 14.88; found: C 57.40, H 9.28, N 14.87.

Benzyl (2S/R,5S,6S)-2-(tert-Butyl)-5-fluorotetrahydro-6-methyl-4-oxopyrimidine-1(2H)-carboxylate (Cbz-11). According to GP 11, 11 (100 mg, 0.53 mmol) was dissolved in CH₂Cl₂ (1.0 ml), and treated with N,O-bis(trimethylsilyl)acetamide (0.20 ml, 0.80 mmol) and Cbz-Cl (0.10 ml, 0.69 mmol). FC (CH₂Cl₂/AcOEt 9:1 → 3:1) yielded Cbz-11 (66.7 mg, 39%). Colorless oil. [α]_D²⁴ = +35.6 (c = 0.77, CHCl₃). IR: 3228w, 2961w, 2911w, 2879w, 1688s, 1483w, 1456w, 1391m, 1318s, 1291s, 1198m, 1160w, 1120w, 1076m, 1038s, 1019m, 967w, 898w, 774m, 736m, 697s. ¹H-NMR (300 MHz, CDCl₃): 1.00 (s, 3 Me); 1.56 (d, J = 6.3, Me); 4.20 – 4.36 (m, NCH); 4.90 (dd, J = 48.2, 8.6, CFH); 5.18 (s, PhCH₂); 5.38 – 5.40 (s, NCHN); 7.10 (s, NH); 7.31 – 7.41 (m, 5 arom. H). ¹³C-NMR (100 MHz, CDCl₃): 20.4 (Me); 26.5 (3 Me); 38.6 (t-Bu); 53.6 (d, J = 29.6, NCH); 68.5 (PhCH₂), 72.9 (NCHN); 86.4 (d, J = 180.1, FCH); 128.2 (arom. C); 128.4 (arom. C); 128.7 (arom. C); 135.6 (arom. C); 152.0 (CO); 167.2 (d, J = 20.2, CFCO). ¹⁹F-NMR (280 MHz, CDCl₃): −195.5 (dd, J = 47.7, 22.6, F). ESI-MS: 323.1758 ([M + H] $^+$, C₁₇H₂₄FN₂O $_3^+$; calc. 323.1765 (err. 2.3 ppm)). Anal. calc. for C₁₇H₂₃FN₂O₃: C 63.34, H 7.19, N 8.69; found: C 63.08, H 7.28, N 8.40

(2S,5R,6S)-2-(tert-*Butyl*)-5-fluorotetrahydro-6-methylpyrimidin-4(1H)-ones (12). According to *GP 10*, aminobutanoic acid 2aca (1.80 g, 7.05 mmol) was converted to the corresponding Cbz-protected amino acid amide 2acd (1.16 g, 65% yield), and subsequent hydrogenation (1.00 g, 3.93 mmol) gave the β-amino acid amide 2aad (467 mg, 99% yield). Amide 2aad (240 mg, 2.00 mmol) was treated with pivalaldehyde to give crude 12 as white solid. Purification by FC (CH₂Cl₂/AcOEt 1:1) afforded 12 (251 mg, 67% yield). Colorless solid. Crystallization gave colorless prisms. M.p. 141–142° (hexane/AcOEt). [α] $_{25}^{26}$ = +116.6 (c = 1.0, CHCl₃). IR: 3321w, 3294w, 3252w, 3193w, 3069w, 2982w, 2950w, 2910w, 2876w, 1671s, 1483m, 1454m, 1411w, 1374m, 1340m, 1287w, 1278w, 1261w, 1234w, 1214w, 1172w, 1144m, 1103w, 1078m, 1049w, 1014w, 993m, 982m, 952m, 938m, 904w, 884w, 869w, 834m, 819s, 798m, 711m, 688m, 661m. ¹H-NMR (400 MHz, CDCl₃): 0.99 (s, 3 Me); 1.29 (dd, J = 6.8, 1.2, Me); 1.42 (br. s, NH); 2.94–3.12 (m, NCH); 4.01 (d, J = 7.0, NCHN); 4.52 (ddd, J = 48.4, 1.8, 0.7, CFH); 6.63 (br. s, NH). ¹³C-NMR (100 MHz, CDCl₃): 15.9 (d, J = 7.9, Me); 25.0 (3 Me); 34.2 (Me₃C); 51.6 (d, d) = 21.0, NCH); 76.2 (NCHN); 87.8 (d, d) = 174.5, FCH); 166.9 (d, d) = 19.1, CFCO). ¹⁹F-NMR (376 MHz, CDCl₃): 46.7 (ddd, d) = 48.5, 30.1, 4.3, 1 F). Anal. calc. for C₉H₁₇FN₂O (188.24): C 57.42, H 9.10, N 14.88; found: C 57.16, H 9.07, N 14.68.

Benzyl (2S/R,5R,6S)-2-(tert-Butyl)-5-fluorotetrahydro-6-methyl-4-oxopyrimidine-1(2H)-carboxylate (Cbz-12). According to GP 11, 12 (100 mg, 0.53 mmol) was dissolved in CH₂Cl₂ (1.0 ml), and treated with N,O-bis(trimethylsilyl)acetamide (0.20 ml, 0.80 mmol) and Cbz-Cl (0.10 ml, 0.69 mmol). FC (CH₂Cl₂/AcOEt 3:1) yielded Cbz-12 (58.1 mg, 34% yield). Crystallization gave colorless prisms. M.p. $124-125^{\circ}$ (Et₂O). $[a]_D^{24} = -62.9$ (c = 1.0, CHCl₃). IR: 3197w, 3121w, 3086w, 2989w, 2961w, 2900w, 1704m, 1677s, 1475w, 1466w, 1458w, 1417m, 1399m, 1390m, 1329m, 1311s, 1299s, 1282s, 1216w, 1193w, 1084s, 1063m, 1046s, 970w, 942w, 899w, 870w, 814w, 774m, 750s, 700s, 630w, 611w. 1 H-NMR (400 MHz, CDCl₃): 1.01 (s, 3 Me); 1.35 (dd, J = 7.2, 2.5, Me); 4.72 (dd, J = 48.1, 6.4, CFH); 5.05 –5.15 (m, NCH); 5.20 –5.21 (m, PhCH₂); 5.39 (s, NCHN); 6.41 (s, NH); 7.35 –7.42 (m, 5 arom. H). 13 C-NMR (100 MHz, CDCl₃): 15.9 (Me); 26.6 (3 Me); 37.3 (Me₃C); 51.7 (d, J = 23.5, NCH); 68.6 (PhCH₂), 72.5 (NCHN); 84.0 (d, J = 192.9, FCH); 128.3 (arom. C); 128.6 (arom. C); 128.7 (arom. C); 135.5 (arom. C); 156.6 (CO); 166.8 (d, J = 20.8, CFCO). 19 F-NMR (376 MHz, CDCl₃): -198.8 (d, J = 48.5, 1 F). ESI-MS: 323.1766 ([M + H]+, C₁₇H₂₄FN₂O₃; calc. 323.1765 (err. -0.2 ppm)); 345.1585 ([M + Na]+, C₁₇H₂₃FN₂NaO₃; calc. 345.1585 (err., 0.0 ppm)). Anal. calc. for C₁₇H₂₃FN₂O₃: C 63.34, H 7.19, N 8.69; found: C 63.09, H 7.12, N 8.65

(2R/S,6S)-2-(tert-Butyl)-5,5-difluorotetrahydro-6-methylpyrimidin-4(1H)-ones (13a/13b). According to GP 10 aminobutanoic acid 3aca (1.40 g, 5.12 mmol) was converted to the corresponding Cbz-protected amino acid amide 3acd (930 mg, 67% yield), and subsequent hydrogenation (800 mg, 2.94 mmol) gave the β -amino acid amide 3aad (405 mg, quant.). Amide 3aad (200 mg, 1.45 mmol) was treated with pivalaldehyde to give crude 13a/13b as white solid. Purification by FC (hexane/AcOEt 7:3,

hexane/acetone 85:15) afforded 13a/13b (73.1 mg, 24%; dr 3:1). Colorless solid. Crystallization gave colorless prisms (1:1 diastereoisomeric co-crystal). M.p. $110-113^\circ$ (hexane/AcOEt). $[a]_D^{20} = -3.1$ (c=1, CHCl₃). IR: 3321w, 3294w, 3252w, 3193w, 3069w, 2982w, 2950w, 2910w, 2876w, 1671s, 1483m, 1454m, 1411w, 1374m, 1340m, 1287w, 1278w, 1261w, 1234w, 1214w, 1172w, 1144m, 1103w, 1078m, 1049w, 1014w, 993m, 982m, 952m, 938m, 904w, 884w, 869w, 834m, 819s, 798m, 711m, 688m, 661m. 1 H-NMR (400 MHz, CDCl₃; *: minor isomer): 0.98 (s, 3 Me); 0.99 (s, 3 Me*); 1.29 (d, J = 5.4, Me*); 1.30 (d, J = 6.6, Me); 1.59-1.65 (m, NH); 2.16-2.24 (m, NH*); 3.10-3.26 (m, NCH); 3.44-3.54 (m, NCH*); 4.08-4.14 (m, NCHN, NCHN*); 6.13 (br. s, NH); 6.24 (br. s, NH). 19 F-NMR (280 MHz, CDCl₃): -104.3 (ddd, J = 277.5, 13.9, 3.9, 1 F*); -120.2 (dd, J = 280.4, 17.9, 1 F); -121.4 (ddd, J = 280.3, 6.6, 2.1, 1 F); -123.8 (d, J = 275.9, 1 F*).

Benzyl (2S/R,6S)-2-(tert-Butyl)-5,5-difluorotetrahydro-6-methyl-4-oxopyrimidine-1(2H)-carboxylate (Cbz-**13**). According to *GP 11*, **13** (150 mg, 0.73 mmol) was dissolved in CH₂Cl₂ (1.4 ml), and treated with *N*,*O*-bis(trimethylsilyl)acetamide (0.27 ml, 1.09 mmol) and Cbz-Cl (0.13 ml, 0.95 mmol). FC (CH₂Cl₂/Et₂O 97:3) yielded Cbz-**13** (15.1 mg, 6%). Colorless oil. IR: 3236w, 2960w, 2875w, 1760w, 1693s, 1483m, 1455w, 1403w, 1373w, 1331w, 1296w, 1284w, 1199s, 1152m, 1104m, 1071m, 1029w, 1003m, 937w, 910w, 805m, 774m, 747m, 697s, 653m. ¹H-NMR (400 MHz, CDCl₃): 1.00 (s, 3 Me); 1.32 (d, J = 6.6, Me); 3.21 (ddq, J = 19.1, 12.7, 6.4, NCH); 4.13 (dd, J = 11.9, 3.4, NCHN); 5.20 (s, PhCH₂); 7.35 – 7.44 (m, 5 arom. H). ¹³C-NMR (100 MHz, CDCl₃): 12.1 (Me); 24.9 (3 Me); 34.2 (Me₃C); 53.3 (t, J = 24.5, NCH); 67.9 (PhCH₂), 75.7 (NCHN); 110.8 (t, J = 247.0, CF₂); 128.1 (arom. C); 128.4 (arom. C); 128.7 (arom. C); 134.9 (arom. C); 151.8 (CO); 163.6 (t, J = 29.8, CFCO). ¹⁹F-NMR (280 MHz, CDCl₃): – 120.5 (dd, J = 282.2, 17.8, 1 F); – 121.7 (ddd, J = 282.2, 6.9, 1.9, 1 F). ESI-MS: 341.1684 ([M + H]+, C₁₇H₂₃F₂N₂O³; calc. 341.1671 (err., – 3.7 ppm)).

5. Preparation of the Fluorinated Cyclo-β-tripeptides **17–19** (Scheme 3, left). 5.1. (S,S)-Isomer **17** from **16a**. Boc-(2S,3S)- $\beta^{2,3}$ -hAla(α -F)-hGly-hGly-OMe (**16a**). According to GP 12a amino fluoro acid **1aba** (700 mg, 3.16 mmol) was converted to the corresponding tripeptide. Purification by FC (hexane/acetone 7:3 \rightarrow 1:1) afforded **16a** (821 mg, 69%). Colorless solid. Crystallization gave colorless prisms. M.p. 155–156° (CH₂Cl₂/Et₂O). [α]₀²⁰ = -22.6 (c=1.0, MeOH). IR: 3350m, 3282m, 3086m, 2977m, 2951m, 1734m, 1689m, 1655m, 1640m, 1554m, 1530m, 1442m, 1387m, 1367m, 1337m, 1311m, 1271m, 1250m, 1171m, 111m, 1062m, 990m, 926m, 883m, 850m, 806m, 781m, 749m, 708m, 643m, 616m. ¹H-NMR (400 MHz, CD₃OD): 1.11 (dd, J = 7.0, 0.5, Me); 1.46 (m, 3 Me); 2.43 (m, m, 6.7, 0.5, CH₂); 2.55 (m, m, 6.6, CH₂); 3.45 (m, J = 6.6, NCH₂); 3.47 – 3.56 (m, NCH₂); 3.70 (m, Me); 4.12 (m, m, 27.3, 10, 3.0, NCH); 4.90 (m, m, 49.7, 3.0, FCH). ¹³C-NMR (100 MHz, CD₃OD): 12.6 (m, m, 4.12 (m, 4) = 27.3, 7.0, 3.0, NCH); 4.90 (m, 4) = 49.7, 3.0, FCH). ¹³C-NMR (100 MHz, CD₃OD): 12.6 (m, 4) = 5.7, Me); 27.3 (3 Me); 33.3 (CH₂); 34.89 (CH₂); 34.93 (CH₂); 35.3 (CH₂); 49.2 (m, m, 20.5, NCH); 50.8 (Me); 79.0 (Me₃C); 92.3 (m, m, 190.6, FCH); 156.0 (CO); 168.8 (m, m, 20.0, CFCO); 172.3 (CO); 172.4 (CO). ¹⁹F-NMR (280 MHz, CD₃OD): -203.7 (m, 4, 4, N 10.99.

Cyclo[(2S,3S)- $β^{2,3}$ -hAla(α-F)-hGly-hGly] (17). According to *GP 3*, 16a (480 mg, 1.27 mmol) was converted to the corresponding carboxylic acid (443 mg, 96% yield). According to *GP 13*, the carboxylic acid (363 mg, 1.00 mmol) was converted to the pentafluorophenyl ester (471 mg, 89% yield), subsequent removal of the Boc group (400 mg, 0.756 mmol) afforded the TFA salt, which was converted (by treatment with 1 Pr₂NEt in MeCN) to 17 (148 mg, 80%). Colorless solid. M.p. 299 – 300° (dec., MeCN). IR: 3388m, 3102w, 2973w, 2949w, 2876w, 1662s, 1646s, 1565s, 1544s, 1454m, 1446m, 1377m, 1351w, 1312w, 1270m, 1237m, 1200m, 1157w, 1133m, 1098m, 1069s, 1029s, 1009m, 988w, 923w, 880w, 864w, 736m, 713m, 671s, 609m. 1 H-NMR (300 MHz, CDCl₃ + TFA): 1.49 (d, d) = 7.4, Me); 2.52 – 2.64 (m, 2 CHH); 2.72 (dt, d) = 14.0, 3.6, 1 H, CH₂); 3.06 (ddd, d) = 14.6, 11.8, 5.6, 1 H, CH₂); 3.29 – 3.44 (m, 2 NCHH); 3.76 (ddd, d) = 14.1, 5.4, 2.6, 1 H, NCH₂); 4.20 – 4.27 (m, 1 H, NCH₂); 4.58 (dq, d) = 21.6, 7.3, NCH); 4.98 (d, d) = 48.6, FCH). 19 F-NMR (280 MHz, CDCl₃ + TFA): – 179.3 (dd, d) = 48.2, 21.2, 1 F). ESI-MS: 246.1241 ([d) + H] ${}^{+}$, C₁₀H₁₆FN₃NaO ${}^{+}$; calc. 246.1248 (err., 3.0 ppm)).

5.2. (R,S)-Isomer **18** from **16b**. $Boc-(2R,3S)-\beta^{2,3}-hAla(\alpha-F)-hGly-hGly-OMe$ (**16b**). According to $GP\ 12a$, amino fluoro acid **2aba** (1.19 g, 5.40 mmol) was converted to the corresponding tripeptide. Purification by FC (hexane/acetone 7:3 \rightarrow 1:1) afforded **16b** (1.51 g, 74%) as colorless solid. Recrystallization gave colorless solid. M.p. $117-118^{\circ}$ (CH₂Cl₂/Et₂O). $[\alpha]_D^{10} = +22.2$ (c=1.0, MeOH). IR: 3336m, 3312m, 3089w, 2973w, 2936w, 1732m, 1686s, 1656s, 1640s, 1526s, 1439m, 1366m, 1354m, 1326m,

1284m, 1247m, 1200m, 1164s, 1104m, 1056m, 1028m, 993m, 921w, 889w, 851w, 782w, 653m. 1 H-NMR (400 MHz, CD₃OD): 1.24 (d, J = 7.0, Me); 1.44 (s, 3 Me); 2.34–2.48 (m, CH₂); 2.56 (t, J = 6.7, CH₂); 3.42–3.56 (m, 2 NCH₂); 3.70 (s, Me); 4.08–4.22 (m, NCH); 4.80 (dd, J = 47.7, 3.0, FCH). 13 C-NMR (100 MHz, CD₃OD): 15.8 (Me); 27.3 (3 Me); 33.3 (CH₂); 35.0 (CH₂); 35.1 (CH₂); 35.4 (CH₂); 47.5 (d, J = 20.4, NCH); 50.8 (Me); 79.0 (Me₃C); 92.5 (d, J = 189.4, FCH); 156.0 (CO); 169.1 (d, J = 21.1, CFCO); 172.3 (CO); 172.5 (CO). 19 F-NMR (376 MHz, CD₃OD): 46.7 (dd, J = 47.7, 24.9, 1 F). Anal. calc. for $C_{16}H_{28}$ FN₃O₆ (377.41): C 50.92, H 7.48, N 11.13; found: C 51.01, H 7.47, N 10.94.

Cyclo[(2R,3S)- $β^{2.3}$ -hAla(α-F)-hGly-hGly] (**18**). According to *GP 3*, **16b** (1.00 g, 2.65 mmol) was converted to the corresponding carboxylic acid (955 mg, 99%). According to *GP 13*, the carboxylic acid (850 mg, 2.34 mmol) was converted to the pentafluorophenyl ester (1.09 g, 88%), subsequent removal of the Boc group (250 mg, 0.472 mmol) afforded the TFA salt, which was converted (by treatment with EtNⁱPr₂ in MeCN) to **18** (74.8 mg, 75%). Colorless solid. M.p. 305° (dec., MeCN). IR: 3329m, 3284m, 3098w, 2971w, 2929w, 2871w, 1670s, 1663s, 1648s, 1550s, 1452m, 1435m, 1371w, 1347m, 1303m, 1275m, 1243m, 1199m, 1187m, 1164w, 1119m, 1094m, 1083m, 1066w, 1056w, 1013m, 991m, 960w, 924w, 886w, 866w, 788w, 738m, 686s, 670m, 632m, 616m. ¹H-NMR (300 MHz, CDCl₃ + TFA): 1.38 (d, J = 7.0, Me); 2.54 (ddd, J = 13.8, 5.3, 1.8, 1 H, CH₂); 2.64 (dt, J = 15.2, 6.2, 1 H, CH₂); 2.82 (ddd, J = 13.8, 12.1, 6.1, 1 H, CH₂); 3.06 (ddd, J = 15.2, 8.1, 5.6, 1 H, CH₂); 3.30 – 3.47 (m, 2 NCHH); 3.70 – 3.79 (m, 1 H, NCH₂); 3.89 – 4.00 (m, 1 H, NCH₂); 4.67 – 4.89 (m, NCH); 5.01 (dd, J = 47.5, 1.6, FCH); 6.81 (d, J = 9.3, NH); 7.35 (br. s, NH); 7.78 (br. s, NH). ¹⁹F-NMR (280 MHz, CDCl₃ + TFA): – 204.4 (ddd, J = 47.3, 32.2, 3.7, 1 F). ESI-MS: 268.1067 ([M + H]⁺, C₁₀H₁₆FN₃NaO₃⁺; calc. 268.1068 (err., 0.5 ppm)).

5.3. Difluoro Derivative **19** from **16c**. Boc-(3S)- $\beta^{2,2,3}$ - $hAla(\alpha,\alpha-F_2)$ -hGly-hGly-OMe (**16c**). According to GP 12a, amino fluoro acid **3aba** (1.00 g, 4.18 mmol) was converted to the corresponding tripeptide. Purification by FC (CH₂Cl₂/acetone 8:2) afforded **16c** (1.21 g, 73%) as colorless solid. Recrystallization gave colorless prisms. M.p. 151–152° (CH₂Cl₂/Et₂O). $[\alpha]_D^{28} = +16.3$ (c=1.0, MeOH). IR: 3345m, 3289m, 3092w, 2975w, 2951w, 1735m, 1693s, 1677m, 1641m, 1533s, 1442m, 1388w, 1367m, 1341m, 1314m, 1269m, 1253m, 1198m, 1172s, 1152m, 1128s, 1105m, 1078m, 1059m, 1044m, 1009m, 997m, 925w, 887w, 870w, 851m, 787w, 754w, 709m, 642m. ¹H-NMR (400 MHz, CD₃OD): 1.21 (d, J = 7.0, Me); 1.45 (s, 3 Me); 2.37–2.50 (m, CH₂); 2.56 (t, J = 6.7, CH₂); 3.44–3.58 (m, 2 NCH₂); 3.69 (s, Me); 4.23–4.35 (m, NCH). ¹³C-NMR (100 MHz, CD₃OD): 12.6 (Me); 27.3 (3 Me); 33.2 (CH₂); 34.7 (CH₂); 34.9 (CH₂); 35.8 (CH₂); 48.3 (t, J = 26.1, NCH); 50.8 (Me); 79.3 (Me₃C); 116.0 (t, J = 255.7, CF₂); 156.0 (CO); 164.2 (t, J = 28.8, CFCO); 172.1 (CO); 172.4 (CO). ¹⁹F-NMR (376 MHz, CD₃OD): -117.5 (dd, J = 251.1, 11.0, 1 F); -119.9 (dd, J = 251.1, 15.1, 1 F). Anal. calc. for C₁₆H₂₇F₂N₃O₆ (395.40): C 48.60, H 6.88, N 10.63; found: C 48.71, H 6.89, N 10.53.

Cyclo[(3S)- $\beta^{2,2,3}$ -*hAla*(α,α - F_2)-*hGly*-*hGly*] (**19**). According to *GP 3*, **16c** (800 mg, 1.84 mmol) was converted to the corresponding carboxylic acid (770 mg, quant.). According to *GP 13*, the carboxylic acid (700 mg, 1.84 mmol) was converted to the pentafluorophenyl ester (880 mg, 87%), subsequent removal of the Boc group (660 mg, 1.21 mmol) afforded the TFA salt, which was converted (by treatment with EtN Pr₂ in MeCN) to **19** (194 mg, 61%). Colorless solid. M.p. 297 – 299° (dec., MeCN). IR: 3307*m*, 3264*m*, 3098*w*, 2986*w*, 2945*w*, 1685*w*, 1664*s*, 1640*s*, 1556*s*, 1538*s*, 1446*m*, 1435*m*, 1372*m*, 1346*w*, 1317*w*, 1278*m*, 1249*m*, 1200*s*, 1185*s*, 1161*m*, 1146*s*, 1118*s*, 1092*m*, 1070*s*, 1023*m*, 1011*m*, 977*m*, 976*w*, 919*w*, 890*w*, 861*w*, 834*w*, 778*w*, 704*m*, 678*s*, 654*s*. ¹H-NMR (300 MHz, CDCl₃ + TFA): 1.36 (*d*, *J* = 6.9, Me); 2.56 (*ddd*, *J* = 14.2, 5.2, 2.0, 1 H, CH₂); 2.62 – 2.87 (*m*, 2 C*H*H); 3.03 (*ddd*, *J* = 15.3, 7.2, 5.4, 1 H, CH₂); 3.40 – 3.57 (*m*, 2 NC*H*H); 3.68 – 3.77 (*m*, 1 H, NCH₂); 3.90 – 4.02 (*m*, 1 H, NCH₂); 4.76 – 4.96 (*m*, NCH); 6.90 (*d*, *J* = 10.2, NH); 7.53 (br. *s*, NH); 7.82 (br. *s*, NH). ¹⁹F-NMR (280 MHz, CDCl₃ + TFA): – 109.9 (*d*, *J* = 256.2, 1 F); – 126.5 (*d*, *J* = 256.2, 21.9, 1 F). ESI-MS: 264.1153 ([*M* + H]⁺, C₁₀H₁₆F₂N₃O⁺₃; calc. 264.1154 (err. 0.6 ppm)).

6. Preparation of the Di- and Tetrafluoro-cyclo- β -tetrapeptides **21–23** (Scheme 3, right). 6.1. (S,S,S,S)-Isomer **21** from Boc-Dipeptide Ester **20a**. Boc-(2S,3S)- $\beta^{2,3}$ -hAla(α -F)-hGly-OMe (**20a**). According to GP 12a, amino fluoro acid **1aba** (2.21 g, 10.0 mmol) was converted to the corresponding dipeptide **20a**. Purification by FC (hexane/AcOEt 65:35 \rightarrow 6:4) afforded **20a** (2.36 g, 77%). Colorless solid. Recrystallization gave colorless prisms. M.p. 114–115° (CH₂Cl₂/Et₂O). [α] $_{0}^{20}$ = -33.0 (c = 1.0, MeOH). IR: 3355m, 2990w, 2942w, 1733s, 1687s, 1657s, 1550m, 1521s, 1446m, 1427w, 1396w, 1367w, 1333m, 1299w, 1268m, 1250s, 1203m, 1182s, 1162s, 1110m, 1084m, 1061s, 993m, 964w, 934w, 880m, 847w,

807w, 779m, 748m, 703m, 639m, 611m. 1 H-NMR (400 MHz, CDCl₃): 1.11 (d, J = 7.0, Me); 1.45 (s, 3 Me); 2.58 (t, J = 6.1, CH₂); 3.54 – 3.65 (m, NCH₂); 3.71 (s, Me); 4.20 – 4.40 (m, NCH); 4.76 (br. s, NH); 4.97 (dd, J = 50.1, 2.2, FCH); 6.88 (br. s, NH). 13 C-NMR (100 MHz, CDCl₃): 14.2 (d, J = 5.3, Me); 28.3 (3 Me); 33.7 (CH₂); 34.4 (CH₂); 47.7 (d, J = 20.4, NCH); 51.9 (Me); 79.8 (Me₃C); 93.0 (d, J = 190.6, FCH); 154.8 (CO); 167.4 (d, J = 19.1, CFCO); 172.6 (CO). 19 F-NMR (376 MHz, CDCl₃): -196.2 (dd, J = 46.5, 18.0, 1 F). Anal. calc. for C₁₃H₂₃FN₂O₅ (306.33): C 50.97, H 7.57, N 9.14; found: C 50.94, H 7.51, N 9.06

Cyclo[(2S,3S)- $β^{2,3}$ -hAla(α-F)-hGly]₂ (21). According to *GP 3*, 20a (1.60 g, 5.22 mmol) was converted to the corresponding carboxylic acid (1.46 g, 95%). According to *GP 13*, the carboxylic acid (1.00 g, 3.42 mmol) was converted to the pentafluorophenyl ester (1.43 mg, 91% yield), subsequent removal of the Boc group (458 mg, 1.00 mmol) afforded the TFA salt, which was converted (by treatment with Pr₂NEt in MeCN) to 21 (203 mg, 58%). Colorless solid. M.p. 249° (dec., MeCN). IR: 3299m, 3093w, 3059w, 2971w, 2961w, 2941w, 1658s, 1561m, 1541s, 1473w, 1449w, 1425m, 1394w, 1380m, 1353w, 1336m, 1276w, 1242w, 1223m, 1203m, 1159m, 1121m, 1083m, 1036m, 999w, 963w, 941w, 923w, 887w, 811w, 769w, 635m. ¹H-NMR (300 MHz, CDCl₃ + TFA): 1.44 (d, J = 7.3, Me); 2.30 (ddd, J = 16.7, 12.0, 3.4, 1 H, CH₂); 2.63 (dt, J = 17.0, 3.1, 1 H, CH₂); 3.42 – 3.53 (m, 1 H, NCH₂); 3.81 – 3.90 (m, 1 H, NCH₂); 4.57 – 4.74 (m, NCH); 4.89 (dd, J = 46.8, 2.1, FCH); 7.00 (dd, J = 9.5, 0.3, NH); 7.75 (br. s, NH). ¹⁹F-NMR (376 MHz, CDCl₃ + TFA): -182.7 (ddd, J = 46.5, 19.5, 3.9, 1 F). ESI-MS: 349.1687 ([M + H]⁺, C₁₄H₂₃F₂N₄O⁴; calc. 349.1682 (err. -1.3 ppm)).

6.2. (R,S,R,S)-Isomer **22** from **20b**. Boc-(2R,3S)- $\beta^{2.3}$ - $hAla(\alpha-F)$ -hGly-OMe **(20b)**. According to GP 12a, amino fluoro acid **2aba** (1.00 g, 4.52 mmol) was converted to **20b**. Purification by FC (hexane/AcOEt 65:35 \rightarrow 6:4) afforded **20b** (1.08 g, 78%). Colorless solid. Recrystallization gave colorless needles. M.p. 82– 83° (CH₂Cl₂/Et₂O). $[a]_D^{28} = +13.9$ (c=1.0, MeOH). IR: 3336m, 3330m, 2981w, 2940w, 1739s, 1682s, 1658s, 1548m, 1522s, 1440m, 1390w, 1367m, 1351m, 1320m, 1267m, 1250s, 1195s, 1164s, 1121w, 1096m, 1073w, 1055m, 1033m, 987m, 942w, 887m, 846m, 814m, 781w, 758w, 742w. ¹H-NMR (400 MHz, CDCl₃): 1.18 (d, J = 6.7, Me); 1.44 (s, 3 Me); 2.58 (dd, J = 6.1, 1.7, CH₂); 3.59 (q, J = 6.0, NCH₂); 3.72 (s, Me); 4.20 – 4.33 (m, NCH); 4.83 (dd, J = 47.1, 1.8, FCH); 5.17 (d, J = 8.6, NH); 6.90 (br. s, NH). ¹³C-NMR (100 MHz, CDCl₃): 16.0 (Me); 28.3 (3 Me); 33.7 (CH₂); 34.4 (CH₂); 47.0 (d, J = 22.3, NCH); 51.9 (Me); 79.6 (Me₃C); 91.6 (d, J = 190.8, FCH); 154.9 (CO); 168.4 (d, J = 19.9, CFCO); 172.6 (CO). ¹⁹F-NMR (376 MHz, CDCl₃): -196.1 (dd, J = 47.1, 18.0, 1 F). Anal. calc. for C₁₃H₂₃FN₂O₅ (306.33): C 50.97, H 7.57, N 9.14; found: C 50.89, H 7.49, N 9.05.

Cyclo[(2S,3S)- $\beta^{2.3}$ -*hAla*(α -*F*)-*hGly*]₂ (**22**). According to *GP 3*, **20b** (1.70 g, 5.55 mmol) was converted to the corresponding carboxylic acid (1.57 g, 97%). According to *GP 13*, the carboxylic acid (1.07 g, 3.66 mmol) was converted to the pentafluorophenyl ester (1.45 g, 86% yield), subsequent removal of the Boc group (458 mg, 1.00 mmol) afforded the TFA salt, which was cyclized by treatment with ${}^{1}\text{Pr}_{2}\text{NEt}$ in MeCN to give **22** (206 mg, 59%). Colorless solid. M.p. 295–296° (dec., MeCN). IR: 3394w, 3325m, 3298m, 3049w, 2974w, 2950w, 2879w, 1654s, 1567m, 1539s, 1444m, 1416m, 1382w, 1349w, 1316m, 1299w, 1283w, 1254m, 1211m, 1166w, 1112m, 1091m, 1079m, 1052w, 1037w, 1024w, 986m, 964m, 924w, 868w, 831w, 771w, 750w, 690m, 667m, 632m, 611m. ${}^{1}\text{H-NMR}$ (300 MHz, CDCl₃+TFA): 1.31 (*d*, *J* = 6.9, Me); 2.46 (*ddd*, *J* = 16.2, 8.4, 3.2, 1 H, CH₂); 2.77 (*ddd*, *J* = 16.2, 8.2, 3.1, 1 H, CH₂); 3.24–3.37 (*m*, 1 H, NCH₂); 3.60–3.74 (*m*, 1 H, NCH₂); 4.13–4.32 (*m*, NCH); 5.05 (*dd*, *J* = 46.7, 1.9, FCH); 7.91 (br. s, NH); 8.12 (*d*, *J* = 8.0, NH). ${}^{1}\text{PF-NMR}$ (280 MHz, CDCl₃+TFA): 40.1 (*dd*, *J* = 46.6, 31.8, 1 F). ESI-MS: 349.1672 ([*M* + H] $^{+}$, C₁₄H₂₃F₂N₄O $^{+}$; calc. 349.1682 (err. – 1.3 ppm)).

6.3. Tetrafluoro-cyclo- β -tetrapeptide **23** from **20c**. Boc-(3S)- $\beta^{2,2,3}$ -hAla(α , α - F_2)-hGly-OMe (**20c**). According to GP 12a, amino fluoro acid **3aba** (1.00 g, 4.18 mmol) was converted to **20c**. Purification by FC (CH₂Cl₂/acetone 8:2) afforded **20c** (952 mg, 71%). Colorless solid. Recrystallization gave colorless prisms. M.p. 115–116° (CH₂Cl₂/Et₂O). [α] $_{\rm D}^{28}$ = +8.7 (c = 1.0, MeOH). IR: 3354m, 2987m, 2955w, 1732m, 1688s, 1671m, 1550m, 1523s, 1446m, 1428w, 1397m, 1368m, 1335m, 1314m, 1253m, 1204m, 1180s, 1157s, 1092w, 1076m, 1056m, 1026m, 1010m, 994m, 927w, 892m, 862w, 845m, 782w, 774w, 740w, 710m, 632m. ¹H-NMR (400 MHz, CD₃OD): 1.21 (d, J = 7.1, Me); 1.45 (s, 3 Me); 2.54–2.66 (m, CH₂); 3.46–3.58 (m, NCH₂); 3.70 (s, Me); 4.30 (ddq, J = 14.6, 11.5, 7.2, FCH). ¹³C-NMR (100 MHz, CD₃OD): 12.6 (Me); 27.3 (3 Me); 32.8 (CH₂); 35.1 (CH₂); 48.2 (t, J = 25.6, NCH); 50.9 (Me); 79.2 (Me₃C); 116.1 (t, J = 255.5, FCH); 156.0 (CO); 164.2 (t, J = 28.8, CFCO); 172.1 (CO). ¹⁹F-NMR (376 MHz, CD₃OD): – 117.5 (dd,

 $J=251.5,\ 11.3,\ 1\ F);\ -119.9\ (dd,\ J=251.5,\ 15.0,\ 1\ F).\ ESI-MS:\ 349.1672\ ([M+H]^+,\ C_{14}H_{23}F_2N_4O_4^+;$ calc. $349.1682\ (err.\ +0.7\ ppm))$. Anal. calc. for $C_{13}H_{22}F_2N_2O_5\ (324.32)$: C 48.14, H 6.84, N 8.64; found: C 48.18, H 6.63, N 8.44.

Cyclo[(3S)- $\beta^{2,2,3}$ -*hAla*(α,α - F_2)-*hGly*]₂ (**23**). According to *GP 3*, **20c** (430 mg, 1.33 mmol) was converted to the corresponding carboxylic acid (406 mg, 97%). According to *GP 13*, the carboxylic acid (400 mg, 1.29 mmol) was converted to the pentafluorophenyl ester (570 mg, 93% yield), subsequent removal of the Boc group (570 mg, 1.20 mmol) afforded the TFA salt, which was cyclized by treatment with EtNⁱPr₂ in MeCN to give **23** (218 mg, 47%). Colorless solid. M.p. 300° (dec., MeCN). IR: 3292m, 3077m, 2989m, 2950m, 1679m, 1651m, 1539m, 1451m, 1415m, 1385m, 1351m, 1305m, 1267m, 1242m, 1198m, 1169m, 1145m, 112m, 1074m, 1006m, 914m, 871m, 840m, 801m, 720m, 705m, 689m. ¹H-NMR (300 MHz, CDCl₃ + TFA): 1.31 (d, d = 6.9, Me); 2.46 (ddd, d = 16.2, 8.4, 3.2, 1 H, CH₂); 2.77 (ddd, d = 16.2, 8.2, 3.1, 1 H, CH₂); 3.24 – 3.37 (m, 1 H, NCH₂); 3.60 – 3.74 (m, 1 H, NCH₂); 4.13 – 4.32 (m, NCH); 5.05 (dd, d = 46.7, 1.9, FCH); 7.91 (br. s, NH); 8.12 (d, d = 8.0, NH). ¹⁹F-NMR (280 MHz, CDCl₃ + TFA): 40.1 (dd, d = 46.6, 31.8, 1 F). ESI-MS: 385.1499 ([d + H]⁺, C₁₄H₂₁F₄N₄O⁺; calc. 385.1493 (err., -1.3 ppm)).

7. Synthesis of the Hexapeptide Derivatives 24 – 26 by Coupling in Solution (Scheme 4). 7.1. The Boc-Hexapeptide Benzyl Ester **24bbb**, Consisting of (S,S)-Residues **1**. Boc-(2S,3S)- $\beta^{2,3}$ -hAla(α -F)-(2S,3S)- $\beta^{2,3}$ $hLeu(\alpha-F)$ -OBn (dipeptide derivative from **1cac** and **1aba**). The benzyl ester **1cbc** (198 mg, 0.56 mmol) was Boc-deprotected according to GP 14a. The resulting TFA salt (1cac · TFA) was dissolved in CH₂Cl₂ (5 ml) and treated with the acid **1aba** (124 mg, 0.56 mmol), NMM (310 μl, 2.8 mmol), HOBt (91 mg, 0.67 mmol), and EDC·HCl (128 mg, 0.67 mmol) according to GP 12b. FC (CH2Cl2/MeOH 100:1) yielded the corresponding dipeptide (173 mg, 68%). Colorless solid. M.p. $148-150^{\circ}$. [a] $_{\rm D}^{\rm r.t}=-40.2$ (c=1.0, CHCl₃). IR (CHCl₃): 3436m, 2964m, 2923w, 2872w, 1759m, 1713s, 1528w, 1503s, 1456m, 1369m, 1333w, 1277w, 1164s, 1128w, 1082w, 1062w, 1031w. 1 H-NMR (400 MHz, CDCl₃): 0.73 (d, J = 6.5, Me); 0.83 (d, J = 6.6, Me); 1.00 (ddd, J = 3.2, 10.2, 13.8, 1 H, CH₂CH); 1.12 (d, J = 6.9, Me); 1.45 (s, t-Bu); 1.45 – 1.58 (m, CHH'CH); 4.28-4.36 (m, NCH); 4.48-4.60 (m, NCH); 4.64 (br. s, BocNH); 4.99 (dd, J=2.3, 50.0, IR)CHF); 5.02 (dd, J = 2.9, 49.1, CHF); 5.16 $(d, J = 11.9, 1 H, PhCH_2)$; 5.35 $(d, J = 11.9, 1 H, PhCH_2)$; 6.36 (br. s, NH); 7.33 – 7.41 (m, 5 arom. H). ¹³C-NMR (100 MHz, CDCl₃): 14.5, 20.7, 23.4 (Me); 24.4 (CH); 28.3 (Me); $36.7 (d, J = 3.3, CH_2)$; 47.7 (d, J = 22.1), 48.3 (d, J = 19.9) (CH); $67.5 (CH_2)$; 79.9 (C); 89.9 (d, J = 19.9)); 47.7 (d, J = 22.1), 48.3 (d, J = 19.9); 47.7 (d, J = 22.1), 48.3 (d, J = 19.9); 47.7 (d, J = 22.1), 48.3 (d, J = 19.9); 47.7 (d, J = 22.1), 48.3 (d, J = 19.9); 47.7 (d, J = 22.1), 48.3 (d, J = 19.9); 47.7 (d, J = 22.1), 48.3 (d, J = 19.9); 47.7 (d, J = 22.1), 48.3 (d, J = 19.9); 47.7 (d, J = 22.1), 48.3 (d, J = 19.9); 47.7 (d, J = 22.1), 48.3 (d, J = 19.9); 47.7 (d, J = 22.1), 48.3 (d, J = 19.9); 47.7 (d, J = 22.1), 48.3 (d, J = 19.9); 47.7 (d, J = 22.1); 48.3 (d, J = 19.9); 47.7 (d, J = 22.1); 48.3 (d, J = 19.9); 47.7 (d, J = 22.1); 48.3 (d, J = 19.9); 47.7 (d, J = 22.1); 48.3 (d, J = 19.9); 47.7 (d, J = 22.1); 48.3 (d, J = 19.9); 47.7 (d, J = 22.1); 48.3 (d, J = 19.9); 47.7 (d, J = 22.1); 48.3 (d, J = 19.9); 47.7 (d, J = 22.1); 48.3 (d, J = 19.9); 47.7 (d, J = 22.1); 47.7 (d, J = 22.1); 48.3 (d, J = 19.9); 47.7 (d, J = 22.1); 47.7 (d, J = 22.1); 48.3 (d, J = 19.9); 47.7 (d, J = 22.1); 47.7 (d, J = 22.1); 47.7 (d, J = 22.1); 48.3 (d, J = 19.9); 47.7 (d, J = 22.1); 47.7 (d, J = 22.1); 48.3 (d, J = 19.9); 47.7 (d, J = 22.1); 47.7 (d, J = 22J = 188.8), 93.1 (d, J = 190.7), 128.7, 128.9, 129.0 (CH); 134.7, 154.7, 167.0 (d, J = 24.1), 167.3 (d, J = 19.2) (C). ¹⁹F-NMR (282 MHz, CDCl₃): -202.8 (dd, J=28.8, 49.1, CHF); -204.6 (dd, J=26.7, 49.1, CHF). HR-MALDI-MS: $495.2 (2, [M+K]^+), 479.2 (61, [M+Na]^+), 423.2 (8), 403.2 (11), 379.2 (32, 12), 423.2 (8), 403.2 (11), 403.2 (12), 403.2 (13), 403.2 (14), 403.2 (15), 403.2 (16), 403.2 (17), 403.2 (18), 4$ $[M + Na - Boc]^+$, 357.2 (100, $[M + H - Boc]^+$), 339.2 (6). Anal. calc. for $C_{23}H_{34}F_2N_2O_5$ (456.53): C 60.51, H 7.51, N 6.14; found: C 60.62, H 7.60, N 6.18.

 $Boc-(2\$,3\$)-\beta^{2,3}-hVal(\alpha-F)-(2\$,3\$)-\beta^{2,3}-hAla(\alpha-F)-(2\$,3\$)-\beta^{2,3}-hLeu(\alpha-F)-OBn$ (**24abb**). The dipeptide derivative (363 mg, 0.79 mmol) described above was Boc-deprotected according to GP 14a. The resulting TFA salt was dissolved in CH₂Cl₂ (4 ml) and treated with the acid **1bba** (200 mg, 0.80 mmol), NMM (440 μl, 4.0 mmol), HOBt (129 mg, 0.95 mmol), and EDC · HCl (183 mg, 0.95 mmol) according to GP 12b. FC (CH₂Cl₂/MeOH 100:5) yielded **24abb** (407 mg, 84%). Colorless solid. R_f (CH₂Cl₂/MeOH 200:3) 0.36. M.p. $189-190^{\circ}$. [α]_{D.} = -49.9 (c=1.0, CHCl₃). IR (CHCl₃): 3432s, 3008w, 2966s, 2874w, 1760m, 1694s, 1522s, 1497s, 1456w, 1392w, 1368m, 1292w, 1161s, 1127m, 1082w, 1030w, 995w, 968w, 903w, 868w, 826w. 1 H-NMR (500 MHz, CDCl₃): 0.73 (d, J = 6.5, Me); 0.84 (d, J = 6.6, Me); 0.93 (dd, J = 0.9, 6.8, Me); 1.00 (d, J = 6.7, Me); 0.99 - 1.06 (m, 1 H, CH₂); 1.19 (d, J = 7.0, Me); 1.44 (s, t-Bu); 1.44 - 1.57 (m, T-Bu); 1.44 - 1.57 (m,1 H, CH₂CH); 1.98 – 2.01 (m, Me₂CH); 3.94 – 4.03 (m, BocNHCH); 4.51 – 4.71 (m, 2 NCH, BocNH); 4.94 (dd, J = 4.1, 48.9, CHF); 4.99 (dd, J = 2.5, 49.8, CHF); 5.02 (dd, J = 3.0, 49.0, CHF); 5.17 (d, J = 11.9, 1 H, 4.10, 1.10, $PhCH_2$; 5.34 (d, $J = 11.9, 1 H, PhCH_2$); 6.42 (br. d, J = 5.8, NH); 6.47 (br. d, J = 4.9, NH); 7.33 – 7.41 (m, 5) arom. H). ¹³C-NMR (125 MHz, CDCl₃): 14.1, 17.9, 20.2, 20.8 (Me); 23.3 (CH); 24.5, 28.3 (Me); 29.6 (d, J = 3.2, CH); 36.8 (CH₂); 46.0 (d, J = 19.5), 48.5 (d, J = 20.0), 56.9 (d, J = 20.9) (CH); 67.5 (CH₂); 79.8 (C); 89.9 (d, J = 188.8), 92.4 (d, J = 191.3), 128.8, 128.9, 129.0 (CH); 134.7, 155.7, 166.9 (d, J = 19.3), 167.0 (d, J=23.8), 167.3 (d, J=19.5) (C). ¹⁹F-NMR (282 MHz, CDCl₃): -193.0 (dd, J=25.6, 49.1, CHF);-202.7 (dd, J = 28.8, 49.1, CHF); -204.2 (dd, J = 26.7, 49.1, CHF). HR-MALDI-MS: 626.3 (3, [M + $K]^+$, 610.3 (32, $[M+Na]^+$), 554.2 (8), 534.2 (13), 510.3 (8, $[M+Na-Boc]^+$), 488.3 (100, $[M+H-Ra]^+$), 610.3 (8, $[M+Na-Rac]^+$), 488.3 (100, $[M+Ra]^+$), 610.3 (8, $[M+Na-Rac]^+$), 488.3 (100, $[M+Rac]^+$), 610.3 (8, $[M+Na-Rac]^+$), 488.3 (100, $[M+Rac]^+$), 610.3 (8, $[M+Rac]^+$), 61 Boc]⁺), 470.2 (8). Anal. calc. for $C_{29}H_{44}F_3N_3O_6$ (587.68): C 59.27, H 7.55, N 7.15; found: C 59.17, H 7.34, N 7.16.

 $Boc-(2\$,3\$)-\beta^{2,3}-hVal(\alpha-F)-(2\$,3\$)-\beta^{2,3}-hAla(\alpha-F)-(2\$,3\$)-\beta^{2,3}-hLeu(\alpha-F)-(2\$,3\$)-\beta^{2,3}-hVal(\alpha-F)-(2\$)-\beta^{2,3}-hVal(\alpha-F)-(2\$)-\beta^{2,3}-hVal(\alpha-F)-(2\$)-\beta^{2,3}-hVal(\alpha-F)-(2\$)-\beta^{2,3}-hVal(\alpha-F)-(2\$)-\beta^{2,3}-hVal(\alpha-F)-(2\$)-\beta^{2,3}-hVal(\alpha-F)-(2\$)-\beta^{2,3}-hVal(\alpha-F)-(2\$)-\beta^{2,3}-hVal(\alpha-F)-(2\$)-\beta^{2,3}-hVal(\alpha-F)-(2\$)-\beta^{2,3}-hVal(\alpha-F)-(2\$)-\beta^{2,3}-hVal(\alpha-F)-(2\$)-\beta^{2,3}-hVal(\alpha-F)-(2\$)-\beta^{2,3}-hVal(\alpha-F)-(2\$)-\beta^{2,3}-hVal(\alpha-F)-(2\$)-\beta^{2,3}-hVal(\alpha-F)-(2\$) (2S,3S)-\beta^{2,3}-hAla(\alpha-F)-(2S,3S)-\beta^{2,3}-hLeu(\alpha-F)-OBn$ (24bbb). Compound 24abb (120 mg, 0.20 mmol) was hydrogenolyzed according to GP 4 to yield the corresponding acid **24aba** (103 mg, quant.). Another sample of 24abb (71 mg, 0.12 µmol) was Boc-deprotected according to GP 14b. The resulting TFA salt of 24aab (60 mg, 0.12 mmol) was dissolved in DMF (3 ml) and treated with 24aba (61 mg, 0.12 mmol), NMM (40 µl, 0.37 mmol), and HATU (56 mg, 0.15 mmol), and the hexapeptide was isolated according to GP 12d. Drying under h.v. yielded **24bbb** (96 mg, 80%). Colorless solid. M.p. > 260° (dec.). ¹H-NMR $(300 \text{ MHz}, (D_6)\text{DMSO}): 0.65 (d, J = 6.5, \text{Me}); 0.75 - 0.90 (m, 7 \text{ Me}); 1.01 - 1.15 (m, 2 \text{ CHH'CH}); 1.02 (d, Me); 0.75 - 0.90 (m, 7 \text{ Me}); 0.75 - 0.90 (m, 7 \text{ Me});$ J = 7.2, Me); 1.04 (d, J = 7.5, Me); 1.43 – 1.49 (m, 2 Me₂CH); 1.66 – 1.82 (m, 2 CHH'CH); 1.84 – 1.94 (m, $2 \text{ Me}_2\text{C}H$); 3.78 - 3.85 (m, BocNHCH); 4.12 - 4.48 (m, 5 NCH); 4.79 (dd, J = 5.9, 48.6, CHF); 4.83 (dd, J = 5.9,J = 2.8, 49.8, CHF; 4.86 (dd, J = 3.1, 49.8, CHF); 4.87 (dd, J = 2.8, 49.5, CHF); 4.94 (dd, J = 5.9, 47.6, 4.9.6, 4CHF); 5.06(dd, J = 3.7, 48.6, CHF); 5.15(d, J = 12.1, 1 H, PhCH₂); 5.26(d, J = 12.1, 1 H, PhCH₂); 6.65(d, J =J = 9.7, NH); 7.36 - 7.40 (m, 5 arom. H); 7.97 (d, J = 8.7, NH); 8.27 (d, J = 7.8, NH); 8.33 (d, J = 8.7, NH); 8.37 (d, J = 8.1, NH); 8.45 (d, J = 8.7, NH). ¹⁹F-NMR (282 MHz, (D₆)DMSO): -192.4 (dd, J = 16.0, 47.0, 4CHF); -193.3 (dd, J = 19.1, 48.0, CHF); -199.4 (dd, J = 26.7, 49.1, CHF); -200.3 (dd, J = 25.6, 49.1, CHF); -201.0 (dd, J = 26.7, 48.0, CHF); -202.1 (dd, J = 27.7, 49.1, CHF). HR-MALDI-MS: 989.5 (100, $[M + Na]^+$, 969.5 (9), 933.5 (8), 913.4 (16), 889.5 (35, $[M + Na - Boc]^+$), 869.5 (41), 867.5 (74, $[M + Na]^+$), 969.5 (9), 938.5 (8), 913.4 (16), 889.5 (35, $[M + Na]^+$), 869.5 (41), 867.5 (74, $[M + Na]^+$), 869.5 (9), 938.5 (9), 913.4 (16), 889.5 (35, $[M + Na]^+$), 869.5 (41), 867.5 (74, $[M + Na]^+$), 869.5 (9), 913.5 (9), 913.4 (16), 889.5 (16), 913.5 $H - Boc^{+}$, 847.5 (30), 827.5 (20), 758.4 (12), 738.4 (5), 586.4 (8).

7.2. The Boc-Hexapeptide Benzyl Ester 25bbb and the Unprotected Hexapeptide 25baa, Consisting of (R,S)-Residues 2. Boc-(2R,3S)- $\beta^{2,3}$ -hAla(α -F)-(2R,3S)- $\beta^{2,3}$ -hLeu(α -F)-OBn (dipeptide derivative from 2cac and 2aba). The benzyl ester 2cbc (431 mg, 1.22 mmol) was Boc-deprotected according to GP 14b. The resulting TFA salt of 2cac was dissolved in CH₂Cl₂ (3 ml) and treated with 2aba (270 mg, 1.22 mmol), NMM (400 µl, 3.7 mmol), and HATU (557 mg, 1.46 mmol) according to GP 12c. FC (hexane/AcOEt 3:1) yielded the corresponding dipeptide (460 mg, 82%). Colorless solid. M.p. 92-93°. $[\alpha]_{\text{b}}^{\text{t.}} = -31.0 \ (c = 1.0, \text{CHCl}_3). \ \text{IR} \ (\text{CHCl}_3): 3429m, 3008w, 2962m, 2923w, 2872w, 1761m, 1707s, 1862w, 1761m, 176$ 1528w, 1501s, 1455m, 1391w, 1368m, 1341m, 1296w, 1165s, 1128w, 1086m, 1044w, 1030w, 847w. ¹H-NMR $(500 \text{ MHz}, \text{CDCl}_3): 0.93 (d, J = 6.7, 2 \text{ Me}); 1.11 (d, J = 6.7, \text{Me}); 1.42 (s, t-Bu); 1.45 - 1.49 (m, CH₂CH);$ 1.55 - 1.63 (m, Me₂CH); 4.15 - 4.28 (m, BocNHCH); 4.64 (qd, J = 8.2, 24.1, NCH); 4.86 (dd, J = 3.5, 47.0, CHF); 4.91 (dd, J = 2.2, 47.2, CHF); 5.12 (br. s, BocNH); 5.15 (d, J = 12.0, 1 H, PhCH₂); 5.23 (d, J = 12.0, 1 H, PhCH₂); 5.23 (d, J = 12.0, 1 H, PhCH₂); 5.24 (d, J = 12.0, 1 H, PhCH₂); 5.25 (d, J = 12.0, 1 H, PhCH₂); 5.26 (d, J = 12.0, 1 H, PhCH₂); 5.27 (d, J = 12.0, 1 H, PhCH₂); 5.28 (d, J = 12.0, 1 H, PhCH₂); 5.29 (d, J = 12.0, 1 H, PhCH₂); 5.20 (d, J = 12.0, 1 H, PhCH₂); 6.20 (d, J = 12.0, 1 H, PhCH₂); 6.2 1 H, PhCH₂); 6.41 (dd, J = 4.2, 9.6, NH); 7.34 – 7.42 (m, 5 arom. H). ¹³C-NMR (125 MHz, CDCl₃): 15.5, $21.9, 22.9 \text{ (Me)}; 24.7 \text{ (CH)}; 28.3 \text{ (Me)}; 39.8 \text{ (CH}_2); 46.8 \text{ } (d, J = 25.5), 48.3 \text{ } (d, J = 21.1) \text{ (CH)}; 67.7 \text{ (CH}_2);$ 79.7 (C); 89.1 (d, J = 189.3), 91.3 (d, J = 192.4), 128.7, 128.8, 128.9 (CH); 134.7, 155.0, 167.5 (d, J = 25.1), 168.0 (d, J = 18.6) (C). ¹⁹F-NMR (282 MHz, CDCl₃): -194.4 (dd, J = 14.9, 45.9, CHF); -230.6 (dd, J = 14.9, CHF24.5, 48.0, CHF). HR-MALDI-MS: 479.2 (46, $[M + Na]^+$), 357.2 (100, $[M + H - Boc]^+$). Anal. calc. for C₂₃H₃₄F₂N₂O₅ (456.53): C 60.51, H 7.51, N 6.14; found: C 60.55, H 7.73, N 6.27.

 $Boc-(2R,3S)-\beta^{2,3}-hVal(\alpha-F)-(2R,3S)-\beta^{2,3}-hAla(\alpha-F)-(2R,3S)-\beta^{2,3}-hLeu(\alpha-F)-OBn$ (25abb). The dipeptide derivative (402 mg, 0.88 mmol) described above was Boc-deprotected according to GP 14b. The resulting TFA salt was dissolved in DMF/CH₂Cl₂ (1:1, 6 ml), and treated with the acid **2bba** (200 mg, 0.80 mmol), NMM (290 µl, 2.64 mmol), and HATU (402 mg, 1.06 mmol) according to GP 12c. FC (CH₂Cl₂/AcOEt 95:5) yielded **25abb** (467 mg, 90%). Colorless solid. R_f (CH₂Cl₂/AcOEt 9:1) 0.22. M.p. $158-159^{\circ}$. [a] $_{1}^{\text{pt}} = -37.1$ (c=1.0, CHCl₃). IR (CHCl₃): 3430m, 3008w, 2966m, 2923w, 2875w, 1761m, 1716s, 1682s, 1521s, 1456w, 1391w, 1368m, 1298w, 1169m, 1135w, 1085m, 1044m, 902w, 863w. ¹H-NMR $(500 \text{ MHz}, \text{CD}_3\text{OD}): 0.89 (d, J = 6.5, \text{Me}); 0.93 (d, J = 6.6, \text{Me}); 0.97 (d, J = 6.7, \text{Me}); 1.04 (d, J = 6.7, \text{Me});$ 1.19 (d, J = 6.9, Me); 1.39 - 1.44 (m, 1 H, CH₂CH); 1.40 (s, t-Bu); 1.60 - 1.65 (m, Me₂CH); 1.67 - 1.72 (m, Me₂CH); 1.67 - 11 H, CH_2CH); 1.85 – 1.92 (m, Me_2CH); 3.80 (ddd, J = 2.0, 8.6, 32.4, BocNHCH); 4.39 – 4.48 (m, NCH); 4.53-4.60 (m, NCH); 4.79 (dd, J = 4.8, 47.6, CHF); 5.02 (dd, J = 2.6, 48.5, CHF); 5.06 (d, J = 12.0, 1 H, $PhCH_2$; 5.18 (dd, J = 2.1, 47.8, CHF); 5.21 (d, J = 12.1, 1 H, $PhCH_2$); 6.48 (d, J = 10.2, NH); 7.31 – 7.44 (m, 5 arom. H). ¹³C-NMR (125 MHz, CD₃OD): 15.8 (*d*, *J* = 2.9), 19.6, 20.0, 21.9, 23.4 (Me); 25.8 (CH); 28.8 (Me); 31.5 (d, J = 2.8, CH); 40.2 (CH₂); 47.3 (d, J = 23.0), 50.1 (d, J = 20.2), 58.8 (d, J = 18.5) (CH); 68.6 (CH_2) ; 80.2 (C); 90.8 (d, J = 187.0), 91.9 (d, J = 186.9), 93.2 (d, J = 191.3), 129.6, 129.6, 129.8 (CH); 136.7, 158.1, 169.5 (d, J = 25.4), 170.1 (d, J = 21.1), 170.5 (d, J = 21.5) (C). ¹⁹F-NMR (282 MHz, CD₃OD): -193.6 (dd, J = 18.1, 48.0, CHF); -202.1 (dd, J = 25.6, 48.0, CHF); -203.3 (dd, J = 32.0, 48.0, CHF). $HR-MALDI-MS: 626.3 (3, [M+K]^+), 610.3 (100, [M+Na]^+), 554.2 (51), 510.3 (22, [M+Na-Boc]^+),$ 488.3 (50, $[M+H-Boc]^+$), 379.2 (7). Anal. calc. for $C_{29}H_{44}F_3N_3O_6$ (587.68): C 59.27, H 7.55, N 7.15; found: C 59.43, H 7.65, N 7.14.

 $Boc-(2R,3S)-\beta^{2,3}-hVal(\alpha-F)-(2R,3S)-\beta^{2,3}-hAla(\alpha-F)-(2R,3S)-\beta^{2,3}-hLeu(\alpha-F)-(2R,3S)-\beta^{2,3}-hVal(\alpha-F)-(2R,3S) (2R,3S)-\beta^{2.3}-hAla(\alpha-F)-(2R,3S)-\beta^{2.3}-hLeu(\alpha-F)-OBn$ (25bbb). Compound 25abb (117.5 mg, 0.2 µmol) was hydrogenolyzed according to GP 4 to yield the corresponding acid 25aba (99.5 mg, quant.). Another sample of 25abb (120 mg, 0.2 μmol) was Boc-deprotected according to GP 14b. The resulting amine 25aab (105 mg, 0.20 mmol) was dissolved in DMF/CH₂Cl₂ 1:1 (3 ml) and treated with 25aba (99.5 mg, 0.20 mmol), NMM (70 μl, 0.60 mmol), and HATU (93 mg, 0.25 mmol), according to GP 12c. FC (CHCl₃/TFE 98:2 \rightarrow 95:5) yielded **25bbb** (118 mg, 60%). Colorless solid. M.p. 234–236°. $R_{\rm f}$ (CHCl₃/ TFE 9:1) 0.25. For ¹H- and ¹³C-NMR spectra, see also Fig. 4. ¹H-NMR (600 MHz, (D₃)TFE/TFA): 0.93 - 1.01 (m, 6 Me); 1.06 (t, J = 7.2, 2 Me); 1.17 (d, J = 6.7, Me); 1.27 (d, J = 6.9, Me); 1.43 (s, t-Bu); 1.44 – 1.63 (m, 2 CH₂CH, 2 Me₂CH); 1.89 – 1.93 (m, Me₂CH); 2.01 – 2.07 (m, Me₂CH); 3.73 – 3.82 (m, BocNHCH); 4.30 (dd, J = 8.2, 30.1, NCH); 4.49 - 4.69 (m, 4 NCH); 4.84 (d, J = 4.2, 46.8, CHF); 4.86 (d, J = 4.2, 46.8,J=3.5, 46.7, CHF); 4.91 (d, J=4.0, 46.3, CHF); 4.98 (d, J=2.8, 47.1, CHF); 5.19 (td, J=2.5, 46.4, 40.5, 40.7, 402 CHF); $5.23 (d, J = 12.1, PhCH_2)$; 7.37 - 7.43 (m, 5 arom. H). ¹³C-NMR (150 MHz, (D₂)TFE/TFA); 15.2, 15.6, 16.0, 19.7, 19.8, 19.9, 20.1, 22.3, 23.3, 23.8 (Me); 26.3, 28.9 (CH); 29.1 (Me); 31.7, 32.2 (CH); 40.3, $41.1 \text{ (CH}_2); 47.7 \text{ } (d, J = 25.6), 48.0 \text{ } (d, J = 23.5), 50.3 \text{ } (d, J = 23.2), 50.9 \text{ } (d, J = 20.6), 57.4 \text{ } (d, J = 18.9), 59.8 \text{ } (d, J = 20.6), 57.4 \text{ } (d, J = 18.9), 59.8 \text{ } (d, J = 20.6), 57.4 \text{ } (d, J = 20.6), 57$ (d, J = 19.0) (CH); 70.2 (CH₂); 82.7 (C); 91.4 (d, J = 187.3), 92.3 (d, J = 190.2), 92.4 (d, J = 189.9), 92.6 (d, J = 190.2), 92.4 (d, J = 189.9), 92.6 (d, J = 190.2), 92.4 (d, J = 189.9), 92.6 (d, J = 190.2), 92.8 (d, J = 190.J = 183.0), 92.7 (d, J = 183.9), 93.3 (d, J = 191.8), 128.8, 129.3, 129.4 (CH); 126.5, 159.1, 170.7 (d, J = 183.9), 93.5 (d, J = 191.8), 128.8, 129.3, 129.4 (CH); 126.5, 159.1, 170.7 (d, J = 183.9), 93.5 (d, J = 191.8), 128.8, 129.3, 129.4 (CH); 126.5, 159.1, 170.7 (d, J = 191.8), 128.8, 129.3, 129.4 (CH); 126.5, 159.1, 170.7 (d, J = 191.8), 128.8, 129.3, 129.4 (CH); 126.5, 159.1, 170.7 (d, J = 191.8), 128.8, 129.3, 129.4 (CH); 126.5, 159.1, 170.7 (d, J = 191.8), 128.8, 129.3, 129.4 (CH); 126.5, 159.1, 170.7 (d, J = 191.8), 128.8, 129.3, 129.4 (CH); 126.5, 159.1, 170.7 (d, J = 191.8), 128.8, 129.3, 129.4 (CH); 126.5, 159.1, 170.7 (d, J = 191.8), 128.8, 129.3, 129.4 (CH); 126.5, 159.1, 170.7 (d, J = 191.8), 128.8, 129.3, 129.4 (CH); 126.5, 159.1, 170.7 (d, J = 191.8), 128.8, 129.4, 129.4 (CH); 126.5, 159.1, 170.7 (d, J = 191.8), 128.8, 129.4, 129.4 (CH); 126.5, 159.1, 170.7 (d, J = 191.8), 128.8, 129.4, 129.4 (CH); 126.5, 159.1, 170.7 (d, J = 191.8), 128.8, 129.4, 129.4 (CH); 126.5, 159.1, 170.7 (d, J = 191.8), 128.8, 129.4, 129.4 (CH); 126.5, 159.1, 170.7 (d, J = 191.8), 128.8, 129.4, 129.4 (CH); 126.5, 159.1, 170.7 (d, J = 191.8), 128.8, 129.4, 129.4 (CH); 126.5, 159.1, 170.7 (d, J = 191.8), 128.8, 129.4, 129.4 (CH); 126.5, 159.1, 170.7 (d, J = 191.8), 128.8, 129.4, 129.4 (CH); 126.5, 159.1, 170.7 (d, J = 191.8), 128.8, 129.4 (CH); 126.5, 159.1, 170.7 (d, J = 191.8), 128.8, 129.4 (CH); 126.5, 159.1, 170.7 (d, J = 191.8), 128.8, 129.4 (CH); 126.5, 159.1, 170.7 (d, J = 191.8), 128.8, 129.4 (CH); 126.5, 159.1, 170.7 (d, J = 191.8), 128.8, 129.4 (CH); 126.5, 159.1, 170.7 (d, J = 191.8), 128.8, 129.8 (d, J = 191.8), 128.8 (d, 20.7), 171.1 (d, J=20.1), 171.2 (d, J=20.9), 171.3 (d, J=25.2), 171.6 (d, J=21.5) (C). ¹⁹F-NMR $(282 \text{ MHz}, (D_6)\text{DMSO}): -190.9 \ (m, 2 \text{ CHF}); -191.5 \ (dd, J = 18.1, 48.0, \text{CHF}); -197.0 \ (dd, J = 26.7, 19.0);$ 47.0, CHF); -198.3 (dd, J = 27.7, 48.0, CHF); -199.8 (dd, J = 25.6, 47.0, CHF). HR-MALDI-MS: 1005.5 $(4, [M+K]^+), 989.5 (75, [M+Na]^+), 889.5 (35, [M+Na-Boc]^+), 867.5 (100, [M+H-Boc]^+), 849.5 (100, [M+H-Boc]^+), 849.5$ (3), 758.4 (5), 488.3 (13).

TFA · *H* · (2R,3S)- $β^{2.3}$ -*hVal*(α-*F*) · (2R,3S)- $β^{2.3}$ -*hAla*(α-*F*) · (2R,3S)- $β^{2.3}$ -*hLeu*(α-*F*) · (2R,3S)- $β^{2.3}$ -*hVal*(α-*F*) · (2R,3S)- $β^{2.3}$ -*hAla*(α-*F*) · (2R,3S)- $β^{2.3}$ -*hLeu*(α-*F*) · (2R,3S)- $β^{2.3}$ -*hLeu*(α-*F*) · (2Sbaa). **25bbb** (35 mg, 36 μmol) was hydrogenolyzed according to *GP* 4 and Boc-deprotected according to *GP* 14a. The crude product was purified by prep. RP-HPLC (MeCN/H₂O + 0.1% TFA 5 : 95 → 60 : 40 (in 25 min) → 90 : 10 (in 5 min) at a flow rate of 18 ml/min) and lyophilized to yield **25baa** (TFA salt; 14.4 mg, 45%). Colorless foam. For ¹H- and ¹³C-NMR spectra, see *Fig.* 4. HR-MALDI-MS: 821 (5), 799 (15, [M + Na]⁺), 777 (100, [M + H]⁺), 759 (5), 543 (3).

7.3. The Boc-Hexapeptide Benzyl Ester 26bbb and the Unprotected Hexapeptide 26baa, Consisting of 2,2-Difluoro-β-amino Acid Residues 3. Boc-(3S)- $\beta^{2,2,3}$ -hAla(α,α - F_2)-(3S)- $\beta^{2,2,3}$ -hLeu(α,α - F_2)-OBn (dipeptide derived from 3cac and 3aba). The benzyl ester 3cbc (500 mg, 1.4 mmol) was Boc-deprotected according to GP 14b. The resulting TFA salt of 3cac (365 mg, 1.35 mmol) was dissolved in DMF (9 ml), and treated with the acid **3aba** (322 mg, 1.35 mmol), NMM (450 µl, 4.04 mmol), and HATU (614 mg, 1.62 mmol) according to GP 12c. FC (hexane/AcOEt $98:2 \rightarrow 9:1$) yielded the corresponding dipeptide (423 mg, 64%). Colorless solid. M.p. 132°. R_f (hexane/AcOEt 4:1) 0.38. $[\alpha]_{\rm L}^{\rm r.t.} = +10.7$ (c=1.0, CHCl₃). IR (CHCl₃): 3426m, 3005w, 2964m, 2933w, 2872w, 1764m, 1713s, 1508s, 1456m, 1364m, 1308w, 1164s, 1092m, 1061m, 1031w, 908w, 861w. ¹H-NMR (500 MHz, CDCl₃): 0.84 (d, J = 6.5, Me); 0.89 (d, J = 6.7, Me); 1.21 (d, J = 7.0, Me); $1.31 - 1.45 (m, CH_2CH)$; 1.44 (s, t-Bu); $1.54 - 1.62 (m, Me_2CH)$; $4.33 - 4.41 (m, CH_2CH)$; NCH); 4.68 (pd, J = 3.1, 11.6, NCH); 4.92 (br. s, BocNH); 5.24 (d, J = 11.9, 1 H, PhCH₂); 5.33 (d, J = 11.9, 1 H, PhCH₂); 5.1 H, PhC H_2); 6.40 (d, J = 10.0, NH); 7.36 – 7.41 (m, 5 arom. H). ¹³C-NMR (125 MHz, CDCl₃): 14.8, 21.0, 23.3 (Me); 24.2 (CH); 28.3 (Me); 36.4 (CH₂); 48.8 (t, J = 26.0), 49.8 (t, J = 24.9) (CH); 68.9 (CH₂); 80.2, 113.9 (t, J = 256.4), 115.8 (t, J = 257.6) (C); 128.8, 128.9, 129.1 (CH); 133.8, 154.9, 162.6 (t, J = 32.2), 163.5 (t, J = 30.0) (C). ¹⁹F-NMR (282 MHz, CD₃OD): -113.0 (dd, J = 12.8, 254.0, 1 F, CF₂); -114.7 (dd, J $12.8, 252.9, 1 \text{ F, CF}_2$; $-115.3 (dd, J = 12.8, 252.9, 1 \text{ F, CF}_2$); $-116.2 (dd, J = 12.8, 252.9, 1 \text{ F, CF}_2)$. HR-MALDI-MS: 531.2 (8, $[M + K]^+$), 515.2 (100, $[M + Na]^+$), 459.2 (76, $[M + Na - isobutylene]^+$), 415.2 $(47, [M + Na - Boc]^+), 393.2 (89, [M + H - Boc]^+)$. Anal. calc. for $C_{23}H_{32}F_4N_2O_5 (492.51)$: C 56.09, H 6.55, N 5.69, F 15.43; found: C 56.17, H 6.68, N 5.61, F 15.42.

 $Boc-(3S)-\beta^{2,2,3}-hVal(\alpha,\alpha-F_2)-(3S)-\beta^{2,2,3}-hAla(\alpha,\alpha-F_2)-(3S)-\beta^{2,2,3}-hLeu(\alpha,\alpha-F_2)-OBn$ (**26abb**). The dipeptide derivative described above (284 mg, 0.57 mmol) was Boc-deprotected according to *GP 14b*. The resulting TFA salt was dissolved in DMF (4 ml), and treated with the acid **3bba** (152 mg, 0.57 mmol),

NMM (190 µl, 1.70 mmol), and HATU (259 mg, 0.68 mmol) according to GP 12c. FC (hexane/AcOEt 95:5 \rightarrow 8:2) yielded **26abb** (270 mg, 73%). Colorless solid. M.p. 122–124°. $R_{\rm f}$ (hexane/AcOEt 4:1) 0.38. $[\alpha]_{\rm D}^{\rm r.t.} = +13.1 \ (c=1.0, {\rm CHCl_3})$. IR (CHCl₃): 3426m, 3261w, 3036w, 2964m, 2875w, 1767s, 1713s, 1525s, 1504s, 1456m, 1392m, 1369m, 1308m, 1158s, 1087m, 1006w, 903w, 872w. ¹H-NMR (500 MHz, CD₃OD): 0.79(d, J = 6.6, Me); 0.89(d, J = 6.7, Me); 0.94(d, J = 6.8, Me); 0.98(d, J = 6.7, Me); 1.22(d, J = 7.0, Me);1.19 - 1.26 (m, 1 H, CH₂CH); 1.43 (s, t-Bu); 1.52 - 1.61 (m, Me₂CH); 1.68 (ddd, J = 3.8, 12.1, 15.7, 1 H, CH_2CH); 2.03 – 2.10 (m, Me_2CH); 4.16 (ddd, J = 4.6, 11.5, 18.7, $CHCF_2$); 4.59 – 4.69 (m, 2 $CHCF_2$); 5.23 $(d, J = 11.9, 1 \text{ H}, PhCH_2)$; 5.34 $(d, J = 11.9, 1 \text{ H}, PhCH_2)$; 7.35 – 7.44 (m, 5 arom. H). ¹³C-NMR (125 MHz, CD_3OD): 13.7, 17.9, 21.0, 21.1, 23.7 (Me); 25.4 (CH); 28.7 (Me); 28.8 (CH); 36.0 (CH₂); 51.3 (t, J = 23.1), 58.1 (t, J = 25.3) (CH); 69.9 (CH₂); 80.7, 115.7 (t, J = 255.5), 117.0 (t, J = 256.1), 118.2 (t, J = 257.1) (C); 129.8, 130.0, 130.1 (CH); 135.9, 158.3, 164.1 (t, J = 32.1), 165.6 (t, J = 28.9), 165.8 (t, J = 29.0) (C). ¹⁹F-NMR (282 MHz, CD_3OD): -110.6 (dd, J = 11.7, 252.9, 1 F, CF_2); -110.8 (dd, J = 9.6, 254.0, 1 F, CF_2); $-113.3 (dd, J = 18.7, 252.9, 1 \text{ F, CF}_2); -113.4 (dd, J = 13.4, 254.0, 1 \text{ F, CF}_2); -115.2 (dd, J = 12.8, 12.8);$ 252.9, 1 F, CF₂); -118.1 (dd, J = 16.6, 252.9, 1 F, CF₂). HR-MALDI-MS: 680.3 (2, $[M + K]^+$), 664.3 (52, $[M+Na]^+$, 608.2 (81, $[M+Na-isobutylene]^+$), 564.2 (54, $[M+Na-Boc]^+$), 542.2 (100, [M+H-isobut]Boc]⁺), 524.2 (6), 466.2 (7), 303.1 (8). Anal. calc. for $C_{29}H_{41}F_6N_3O_6$ (641.65): C 54.28, H 6.44, N 6.55, F 17.77; found: C 54.08, H 6.57, N 6.56, F 17.75.

 F_2)-(3S)- $\beta^{2,2,3}$ - $hAla(\alpha,\alpha-F_2)$ -(3S)- $\beta^{2,2,3}$ - $hLeu(\alpha,\alpha-F_2)$ -OBn (26bbb). Compound 26abb (145 mg, 0.226 µmol) was C-terminally deprotected by hydrogenolysis according to GP 4 to yield the corresponding acid 26aba (107 mg, 86%). Another sample of 26abb (125 mg, 0.194 µmol) was Bocdeprotected according to GP 14b. The resulting TFA salt of 26aab (105 mg, 0.194 mmol) was dissolved in DMF (3 ml), and treated with the acid 26aba (107 mg, 0.194 mmol), NMM (57 µl, 0.52 mmol), and HATU (88 mg, 0.23 mmol) according to GP 12c. The crude product was purified by prep. RP-HPLC $(MeCN/H_2O + 0.1\% TFA 10:90 \rightarrow 55:45 (in 2 min) \rightarrow 67:33 (in 23 min) \rightarrow 90:10 (in 2 min) at a flow$ rate of 18 ml/min) and lyophilized to yield 26bbb (72 mg, 38%). Colorless solid. M.p. 208-210°. 19F-NMR (282 MHz, CD_3OD): -109.1 (dd, J=13.9, 257.2, CFF'); -109.3 (dd, J=8.5, 254.0, 1 F, CF_2); $-110.6 (dd, J = 11.7, 252.9, 1 \text{ F, CF}_2); -110.7 (dd, J = 13.9, 257.2, 1 \text{ F, CF}_2); -111.2 (dd, J = 9.6, 252.9, 1.1)$ 1 F, CF_2 ; $-111.9 (dd, J = 10.7, 254.0, 1 \text{ F, CF}_2)$; $-112.4 (dd, J = 14.9, 256.1, 1 \text{ F, CF}_2)$; $-113.3 (dd, J = 14.9, 256.1, 1 \text{ F, CF}_2)$; -113.312.8, 252.9, 1 F, CF₂); -115.3 (dd, J = 12.8, 254.0, 1 F, CF₂); -117.0 (dd, J = 17.1, 252.9, 1 F, CF₂); -117.0117.1 $(dd, J = 16.0, 255.0, 1 \text{ F, CF}_2)$; $-117.6 (dd, J = 16.0, 252.9, 1 \text{ F, CF}_2)$. HR-MALDI-MS: 1097 (6, $[M + Na]^+$, 1041 (28, $[M + Na - isobutylene]^+$), 1013 (13), 997 (100, $[M + Na - Boc]^+$), 975 (6, $[M + Na]^+$), 1041 (28, $[M + Na]^+$), 1041 (28, $[M + Na]^+$), 1041 (10, $[M + Na]^+$), 1041 (11, $[M + Na]^+$), 10 $H - Boc]^+$, 957 (15), 921 (8), 848 (6).

 $TFA \cdot H \cdot (3S) \cdot \beta^{2,2,3} - hVal(\alpha,\alpha-F_2) \cdot (3S) \cdot \beta^{2,2,3} - hAla(\alpha,\alpha-F_2) \cdot (3S) \cdot \beta^{2,2,3} - hLeu(\alpha,\alpha-F_2) \cdot (3S) \cdot \beta^{2,2,3} - hAla(\alpha,\alpha-F_2) \cdot (3S) \cdot (3S)$

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